- 1 Systematic Entomology, School of Agriculture, Hokkaido University, Sapporo 060-8589,
- 2 Japan; 2 Illinois Natural History Survey, University of Illinois, Champaign, Illinois 61820,
- 3 USA

4

- 5 Wing base structure supports Coleorrhyncha + Auchenorrhyncha (Insecta:
- 6 **Hemiptera**)

7

8 Kazunori Yoshizawa 1, Naoki Ogawa 1 and Christopher H. Dietrich 2

- 10 Corresponding author: Kazunori Yoshizawa (psocid@res.agr.hokudai.ac.jp)
- 11 Contributing authors: Naoki Ogawa (ogawa 222@res.agr.hokudai.ac.jp), Christopher H.
- 12 Dietrich (chdietri@illinois.edu)

Abstract

14

13

The phylogenetic placement of the moss bugs (Insecta: Hemiptera: Coleorrhyncha) has been 15 highly controversial. Many apparent morphological apomorphies support the close 16 17 relationship between Coleorrhyncha and Heteroptera (= true bugs). However, a recent phylogenomic study strongly supported a sister-group relationship between Coleorrhyncha 18 and Auchenorrhyncha (planthoppers, leafhoppers, treehoppers, spittlebugs and cicadas). To 19 test these two alternative hypotheses, we examined the fore- and hindwing base structure of 20 the only known extant macropterous species of Coleorrhyncha using binocular and confocal 21 laser scanning microscopes and analyzed the data selected from the wing base 22 phylogenetically. When full morphological data including the wing base characters were 23 analyzed, the sister group relationship between Coleorrhyncha + Heteroptera was supported, 24 agreeing with previous consensus based on morphology. In contrast, when only wing base 25 characters were analyzed separately, the clade Coleorrhyncha + Auchenorrhyncha was 26 recovered, in agreement with the result from the phylogenomic study. The membranous 27 28 condition of the proximal median plate in the forewing was identified as a potential synapomorphy of the latter grouping, and absence of the tegula was excluded as a potential 29 synapomorphy of Coleorrhyncha and Heteroptera. 30

- **Key words:** Auchenorrhyncha Coleorrhyncha Heteroptera phylogeny wing base
- 33 structure

Introduction

1	_
ń	`

36	The suborder Coleorrhyncha (moss bugs) is an enigmatic taxon of the order
37	Hemiptera (Insecta). It consists of a single family, Peloridiidae, with fewer than 40 extant
38	species restricted to circumantarctic regions (Burckhardt 2009; Burckhardt et al. 2011). With
39	a combination of plesiomorphic and apomorphic features, the placement of this suborder
40	within Hemiptera had been highly unstable. Traditionally, prior to explicit phylogenetic
41	analyses of Hemiptera as a whole, Coleorrhyncha was regarded as a member of "Homoptera"
42	(now generally regarded as a paraphyletic grade) due to presence of a complete tentorium,
43	origin of the labium on the posteroventral portion of the head (and absence of a gula),
44	discrete pro- and mesothracic ganglia, and eight pairs of abdominal spiracles, all of which are
45	now regarded as plesiomorphies (Carver et al. 1991). In contrast, Coleorrhyncha share some
46	apparent morphological apomorphies with Heteroptera (reviewed in Grimaldi and Engel
47	2005; Forero 2008; Burckhardt 2009), some of which have been controversial (e.g., Cobben
48	1978 but see also Schuh 1979). Recent extensive morphology-based cladistic analysis, with
49	revised morphological observations including Coleorrhyncha, strongly supported
50	Coleorrhyncha + Heteroptera (Friedemann et al. 2014). Multiple molecular phylogenetic
51	studies based on 18S rRNA (Wheeler et al. 1993; Campbell et al. 1995; Ouvrard et al. 2000)
52	and multiple gene regions (Cryan and Urban 2012) also provided support for this relationship.
53	Therefore, until recently, available data appear to have converged toward consensus in
54	support of the sister group relationship between Coleorrhyncha and Heteroptera (together
55	referred to as Heteropterodea or Prosorrhyncha) (Grimaldi and Engel 2005; Forero 2008).
56	However, a recent phylogenomic study of Hexapoda that incorporated data from
57	>1400 gene regions (Misof et al. 2014) casted doubt on this general view, placing
58	Coleorrhyncha consistently as sister to Auchenorrhyncha (infraorder composed of
59	planthoppers, leafhoppers, treehoppers, spittlebugs and cicadas). This result was supported by
60	multiple datasets (i.e., nucleotide and amino acid sequences) and also received strong
61	statistical support by bootstrapping and four-cluster likelihood mapping analyses (Misof et al.
62	2014, Supplement). The previously accepted sister group relationship between Coleorrhyncha
63	and Heteroptera was also refuted by recent mitochondrial phylogenomic analyses (Cui et al.
64	2013; Wang et al. 2015). Therefore, an apparent conflict between morphological and
65	molecular data has arisen in the placement of Coleorrhyncha.

The wing base structure comprises sclerites located between the insect thorax and wing. This structure mediates the power produced by the thoracic indirect flight muscles to the wings and also controls proper flapping and folding of the wings. Therefore, evolution of this structure is strongly constrained and, thus, the wing base sclerites appear to evolve very slowly (Hörnschemeyer 2002). Because of this unique property, the wing base structure has previously been utilized for resolving controversial branches in hemipteroid phylogeny. For example, although the monophyly of Auchenorrhyncha has been questioned based on morphological (Bourgoin 1986ab 1993; Bourgoin and Huang 1990) and molecular criteria (Campbell et al. 1995; Sorensen et al. 1995; Bourgoin et al. 1997; Ouvrard et al. 2000), examination of wing base morphology provided unambiguous support for Auchenorrhyncha (Yoshizawa and Saigusa 2001). Monophyly of Auchenorrhyncha subsequently received strong support from the molecular phylogenetic (Urban and Cryan 2007; Cryan and Urban 2012) and phylogenomic (Misof et al. 2014) analyses, corroborating the value of wing base structure for resolving difficult higher-level phylogenetic problems (see also Yoshizawa 2011).

In this study, we examined the morphology of the fore- and hindwing base structures of a species of Coleorrhyncha, which were treated as missing characters by Friedemann et al. (2014), to test the alternative hypotheses on the phylogenetic placement of this suborder.

Material and Methods

A dried specimen of *Peloridium hammoniorum* Breddin, 1897 collected in Chile in 2014 by CHD was used. This is the only extant species of Coleorrhyncha known to have flight ability, although most individuals of this species have vestigial hindwings. The individual studied possessed fully developed fore- and hindwings. The specimen was soaked with 10% KOH at room temperature for one night. Later, the pterothorax was separated from the other body parts, washed by distilled water, then 80% ethanol, and finally stored and observed in glycerol.

Observations were made with an Olympus SZX 16 binocular microscope (Olympus Co., Tokyo, Japan) and Leica TCS-SP5 Confocal Laser Scanning Microscope (CLSM) (Leica Microsystems, Wetzlar, Germany). For binocular microscope observation, the dissected specimen was pinned on a polyfoam using micro-pins, with the wings fully opened

but oriented in a slight downstroke position to observe all the sclerites in their natural shapes dorsally. For CLSM imaging, specimens were mounted on a large cover glass (22 x 24 mm) covered by a small cover glass (15 mm ø) to facilitate the observation of both dorsal and ventral sides. We used an excitation wavelength of 488 nm and emission wavelength of 510–680 nm. The emission waves were detected using two channels and visualized with two pseudocolors (510–580 nm in green; 580–680 nm in red) (Mikó and Deans 2014). Homology was assessed following Yoshizawa and Saigusa (2001), in which the criteria and landmarks for homology identification of paraneopteran (hemipteroid) wing base sclerites were explained. Terminology of Yoshizawa and Saigusa (2001) was also adopted.

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

Morphological data selected from the forewing base of *Peloridium* were newly appended to two data matrices created previously: (1) the forewing base character matrix for Paraneoptera, comprising 20 discrete characters, constructed by Yoshizawa and Saigusa (2001), in which Coleorrhyncha was not examined; (2) the full morphological data, comprising 119 characters (including the above as characters 20–39) compiled by Friedemann et al. (2014), with some corrections to character coding as mentioned by Yoshizawa and Lienhard (2016). The genus *Hackeriella* was used in the original full morphological data matrix (Friedemann et al. 2014) but, because this genus lacks flight ability, almost all wing base characters were previously coded as unknown. Here, the same set of wing base characters scored for the separate wing base matrix was newly appended to the matrix. Although this combination of data from two different peloridiid species created a chimeric OTU in the data matrix, members of the family appear to be invariant for most (if not all) of the included characters, so we would not anticipate a different phylogenetic result had we scored all of Friedemann's characters for Peloridium. The hindwing base structure was also observed but not included in the phylogenetic analyses to avoid the possibility of over-weighting serially homologous (non-independent) traints, as discussed by Yoshizawa and Saigusa (2001). Data matrices are available as online Supporting Information. The datasets were analyzed by the maximum parsimony method using PAUP* 4a151 (Swofford 2002), with all characters weighted equally and branch-and-bound search performed. The branch-and-bound method uses an exact algorithm that is guaranteed to find the most parsimonious tree(s). Bootstrap and jackknife values were calculated using PAUP* with 1000 replicates. For bootstrapping and jackknifing, heuristic searches with tree-bisection-reconnection algorithm were performed, each with 100 replications and with

unlimited maxtrees. The decay index was calculated by using TreeRot V3 (Sorenson and Franzosa 2007). The character state changes were calculated using MacClade 4 (Maddison and Maddison 2001), and unambiguous character state changes were mapped onto the tree.

133

130

131

132

Results

134135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

Forewing base morphology (Fig. 1)

The anterior and median notal wing processes (ANWP and MNWP) are easily identified as articular points with 1Ax: the former is well projecting but the latter is not. The posterior notal wing process (PNWP) is less recognizable because it does not project and is loosely associated with the third axillary sclerite. The tegula (Tg) is apparently present but weakly sclerotized. The humeral plate (HP) is united with the basisubcostale (BSc). The basiradiale (BR) is broadly united with BSc anteriorly, and is united with the second axillary sclerite (2Ax) posteroproximally. The first axillary sclerite (1Ax) is subtriangular in shape (see Fig. 1A; 1Ax is oriented laterally in Fig. 1B), lacking the anterior arm, with a weak swelling on the proximal margin which articulates with MNWP. The anterior tip of 1Ax articulates with the tip of BSc. The posteroproximal margin of 1Ax is also associated with the notum. Distally, 1Ax articulates with 2Ax at two points; near the anterior tip and near the posterodistal corner. 2Ax is subdivided into two sclerites, posteroproximal (pp) and anterodistal (ad), clearly divided by a membranous region and the forked convex axillary fold line. 2Ax-pp is tightly united with the apex of BSc anteroproximally and is articulated with the well-developed and narrowly extended anterior arm of the third axillary sclerite (3Ax) at its distal end. 2Ax-ad is united distally with the first distal median plate, and the convex axillary fold line forms a border between the two sclerites. The proximal arm of 3Ax is broadened and loosely articulated with the notum. Distally, it is tightly associated with the basianal (BA) along the posterior margin (see Fig. 1A,C: in Fig. 1B, 3Ax and BA are detached but this was caused artificially by slide mounting pressure). BA is well developed, tightly articulated with the anal vein distally. The region corresponding to the proxomal median plate (PMP) is completely membranous (see Fig. 1C). The distal median plate (DMP) is subdivided into two elements. DMP1 is enlarged and convex dorsally. It is united proximally with 2Ax, tightly associated with vein R anteriorly, and tightly articulated with vein A posterodistally. DMP2 is located distal to DMP1, triangular in shape and very

narrowly extending toward the claval fold. The jugum (Jg) is apparently not developed.

Hindwing base morphology (Fig. 2)

[Note for CLSM image (Fig. 2B): Due to the less tight articulation of the hindwing sclerites and pressure caused during slide mounting, the positions of many sclerites in the CLSM image are distorted. See the line drawing (Fig. 2A) for their more natural articular condition.]

ANWP and MNWP are recognizable but very loosely articulated with 1Ax, with the former located anterior to the tip of 1Ax. PNWP is well developed and articulated with 3Ax. Tg is absent. HP is united with BSc. BR is only recognizable as a small projection extending from the posterior margin of BSc, and loosely associates with 2Ax at the tip. 1Ax is narrowed over almost its entire length, only weakly broadened medially along the distal margin, with a weakly developed anterior arm. The anterior tip of 1Ax only weakly articulates with the tip of BSc. Distally, 1Ax articulates with 2Ax only at one point. 2Ax is not subdivided as in the forewing but reduced in size and triangular in shape. It tightly articulates with 1Ax only at its anteroproximal corner and also tightly articulates with 3Ax at its posterior tip. 3Ax is well developed, rather loosely articulated with PNWP. Distally, it is tightly associated with the anal vein (distal margin) and DMP (anterodistal corner). BA is indistinguishable from 3Ax. The region corresponding to PMP is completely membranous. DMP is flat, trapezoidal in shape. Jg is large but only weakly sclerotized.

Phylogenetic analyses

The parsimony analysis of the forewing base dataset resulted in only one most parsimonious tree (Fig. 3: treelength = 23; consistency index = 0.91; retention index = 0.93). This tree is completely congruent with that estimated by Yoshizawa and Saigusa (2001), with monophyly of Paraneoptera, Condylognatha, and Hemiptera all supported. Coleorrhyncha (excluded from the analysis of Yoshizawa and Saigusa 2001) formed a clade together with the auchenorrhynchous infraorders (Cicadomorpha and Fulgoromorpha), supported by one unique, non-homoplasious synapomorphy (decay index = 1): the membranous proximal median plate (Character 13:1).

The parsimony analysis of the full morphology dataset, including the forewing base characters, resulted in 18 equally parsimonious trees (treelength = 197; consistency index =

0.69; retention index = 0.85). Fig. 4 shows the strict consensus of 18 trees (differences between them mostly concern the arrangements of zero-length branches and do not affect to the following discussion: see Supporting Information for all trees). The tree is congruent with that estimated by Friedemann et al. (2014), with Paraneoptera, Psocodea, Condylognatha, Hemiptera, Auchenorrhyncha and Heteroptera supported as monophyletic. Coleorrhyncha was placed as sister of Heteroptera, with three apomorphies (including two non-homoplasious ones) supporting this placement. One of the characters selected from the wing base (Character 28:1) provided non-homoplasious support for Hemiptera. The character providing support for Auchenorrhyncha + Coleorrhyncha in the wing base dataset (Character 13:1 in the wing base matrix; 32:1 in the full data matrix) was only ambiguously reconstructed: i.e., either independently evolved between Auchenorrhyncha and Coleorrhyncha or gained in the common ancestor of Auchenorrhyncha + Coleorrhyncha + Heteroptera but reversed in Heteroptera.

Discussion

The wing base structure in Coleorrhyncha largely retains the groundplan condition of the neopteran wing base, except for the absence of a proximal median plate (Figs 1–2). All modifications detected previously and thought to be autapomorphic for Hemiptera in general (Yoshizawa and Saigusa 2001) were also observed in Coleorrhyncha. Overall, the wing base structure of Coleorrhyncha resembles that of Auchenorrhyncha rather than Heteroptera (Yoshizawa and Saigusa 2001; Yoshizawa and Wagatsuma 2012; Ogawa et al. 2015). Maximum parsimony analysis of the wing base characters alone clearly supported the monophyly of Coleorrhyncha + Auchenorrhyncha, with absence of PMP as a synapomorphy (Fig. 3: decay index = 1, bootstrap/jackknife values = 67/100%). As mentioned by Yoshizawa and Saigusa (2001), this is a "reduction" character, i.e., presumably resulting from the loss of a sclerite, which may be regarded as less reliable than a character "gain". However, this character state was previously regarded as an autapomorphy of Auchenorrhyncha, a group once thought to be paraphyletic based on early single-gene molecular phylogenies (e.g., Campbell et al. 1995) but more recently supported as monophyletic by multi-gene molecular phylogenies (Urban and Cryan 2007; Cryan and Urban 2012; Misof et al. 2014). In addition, the present examination clearly identified the tegula on the forewing of Coleorrhyncha, which

invalidates "absence of tegula" as one of the previously proposed synapomorphies of Coleorrhyncha + Heteroptera (Friedemann et al. 2014).

Nevertheless, analysis of the full morphological data still recovered the sister group relationship between the Coleorrhyncha and Heteroptera (Fig 4: Friedemann et al. 2014), although with lower support values (decay index = 1, bootstrap/jackknife values = 52/66%). Synapomorphies supporting this relationship include presence of cephalic trichobothria (54-1), tubular and four-segmented labium (56-2), and four-segmented flagellomeres (59-1). The first two are non-homoplasious characters (Friedemann et al. 2014) in the present dataset. The full morphological matrix constructed by Friedemann et al. (2014) lacked some morphological characters previously suggested as additional synapomorphies of Coleorrhyncha + Heteroptera (Grimaldi and Engel 2005; Forero 2008; Burckhardt 2009; Spangenberg et al. 2013) so it is possible that morphological support for the monophyly of this group is stronger than shown in our analysis.

In contrast, morphological support for the Coleorrhyncha + Heteroptera may not be as robust as generally assumed. For example, the position of abdominal spiracle 2 on an epipleurite was previously suggested as a potential synapomorphy of Coleorrhyncha and Auchenorrhyncha (Sweet 1996). However, because almost all other morphological characters supported a closer relationship between Coleorrhyncha and Heteroptera, Sweet (1996) concluded that the spiracle condition was independently gained by Coleorrhyncha and Auchenorrhyncha. The cephalic trichobothria (54-1) were scored as present for Coleorrhyncha and Heteroptera (Friedmann et al. 2014) and identified as one of their non-homoplasious synapomorphies (Fig. 3). However, the cephalic trichobothria were not illustrated or reported in the recent detailed study of the adult head of Hackeriella (Spangenberg et al. 2013) so the status of this character as a synapomorphy of Coleorrhyncha + Heteroptera is questionable. Spangenberg et al. (2013) further reviewed morphological evidence supporting the monophyly of Coleorrhyncha + Heteroptera but pointed out that the homologies of some potential synapomorphies (e.g., the number of antennomeres) remain uncertain while others (e.g., absence of cervical sclerites) are homoplasious. They also noted several potential synapomorphies of Coleorrhyncha and Auchenorrhyncha, or Coleorrhyncha and "Homoptera" in general. Some of the latter, including absence of a gula and presence of a complete tentorium, were interpreted as plesiomorphic for Hemiptera as a whole, but presence of Evans' organ (Bourgoin 1986b) may be another unique synapomorphy of

258	Coleorrhyncha and Auchenorrhyncha or an autapomorphy of "Homoptera" (including
259	Coleorrhyncha). Characters of the cephalic musculature were found that support either
260	Coleorrhyncha + Heteroptera or Coleorrhyncha + "Homoptera" (Spangenberg et al. 2013).
261	The results from recent phylogenomic analyses (Misof et al. 2014) and the present
262	wing base examination suggest that some of the above-mentioned morphological similarities
263	between Coleorrhyncha and Auchenorrhycha may be their true synapomorphies. In addition,
264	some of the features previously interpreted as synapomorphies of Coleorrhyncha and
265	Heteroptera (Schlee 1969) have already been considered as "superficial and probably not
266	significant" (Cobben 1978: but see also Schuh 1979 for critique). Based on our study, we also
267	exclude "absence of the tegula" (20-1) as a synapomorphy of Coleorrhyncha + Heteroptera.
268	This resulted in a decrease in branch support for this clade from decay index of three
269	(Friedemann et al. 2014) to only one (Fig. 4). Further morphological investigations, including
270	re-evaluation of the previously proposed synapomorphies of Coleorrhyncha and Heteroptera
271	and incorporation of these and various cephalic characters mentioned by Spangenberg et al.
272	(2013) into an explicit phylogenetic analysis, are needed to elucidate the extent of conflict
273	between morphology and phylogenomics and between different morphological character
274	systems.
275	
276	Acknowledgments
277	This study was partly supported by Japan Society for the Promotion of Science pre-doctoral
278	fellowship program (15J03697) to NO and U.S. National Science Foundation grant
279	DEB-1239788 to CHD.
280	
281	References
282	
283	Bourgoin T (1986a) Morphologie imaginale du tentorium des Hemiptera Fulgoromorpha. Int
284	J Ins Morph Embryo 15 :237–252.
285	Bourgoin T (1986b) Valeur morphologique de la lame maxillaire chez les Hemiptera;
286	remarques phylogénétiques. Ann Soc Entomol France (NS) 22:413-422.
287	Bourgoin T (1993) Female genitalia in Hemiptera Fulgoromorpha, morphological and
288	phylogenetic data. Ann Soc Entomol France (NS) 29:225–244.
289	Bourgoin T, Huang J (1990) Morphologie comparée des genitalia mâles des

290	Trypertimorphini et remarques phylogénétiques (Hemiptera: Fulgoromoprha:
291	Tropiduchidae). Ann Soc Entomol France (NS) 26:555-564.
292	Bourgoin T, Steffen-Campbell JD, Campbell BC (1997) Molecular phylogeny of
293	Fulgoromorpha (Insecta, Hemiptera, Archaeorrhyncha). The Enigmatic
294	Tettigometridae: Evolutionary Affiliations and Historical Biogeography. Cladistics
295	13 :207–224.
296	Breddin G (1897) Hemipteren (Hemiptera). Ergebnisse der Hamburger magalhaensischen
297	Sammelreise, 1892/93, vol. 2. L. Friederichsen and Co., Humburg.
298	Burckhardt D (2009) Taxonomy and phylogeny of the Gondowanan moss bugs or
299	Peloridiidae (Hemiptera, Coleorrhyncha). Dtsch Entomol Ztg 56:173–235.
300	Burckhardt D, Bochud E, Damgaard J, Bibbs GW, Hartung V, Larivière MC, Wyniger D,
301	Zürcher I (2011) A review of the moss bug genus Xenophyes (Hemiptera:
302	Coleorrhyncha: Peloridiidae) from New Zealand: systematics and biogeography.
303	Zootaxa 2923 :1–26.
304	Campbell BC, Steffen-Campbell JD, Sorensen JT, Gill RJ (1995) Paraphyly of Homoptera
305	and Auchenorrhyncha inferred from 18S rDNA nucleotide sequences. Syst Entomol
306	20 :175–194.
307	Carver M, Gross GG, Woodward TE (1991) Hemiptera (Bugs, leafhoppers, cicadas, aphids,
308	scale insects etc). In: CSIRO (ed), The Insects of Australia. 2nd edition. Cornell
309	University Press, New York, pp 429–509.
310	Cobben RH (1978) Evolutionary trends in Heteroptera Part II. Mouthpart-structures and
311	feeding strategies. Mededelingen Landbouwhogeschool Wageningen 78(5):1-407.
312	Cryan JR, Urban JM (2012) Higher-level phylogeny of the insect order Hemiptera: is
313	Auchenorrhyncha really paraphyletic? Syst Entomol 37:7–21.
314	Cui Y, Xie Q, Hua J, Dang K, Zou J, Liu X, Wang G, Yu X, Bu W (2013) Phylogenomics of
315	Hemiptera (Insecta: Paraneoptera) based on mitochondrial genomes. Syst Entomol
316	38 :233–245.
317	Forero D (2008) The systematics of the Hemiptera. Rev Columbiana Entomol 34:1–21.
318	Friedemann K, Spangenberg R, Yoshizawa K, Beutel RG (2014) Evolution of attachment
319	structure in the highly diverse Acercaria (Hexapoda). Cladistics 30:170-201.
320	Grimaldi D, Engel MS (2005) Evolution of the Insects. Cambridge Univ. Press, Cambridge.
321	Hörnschemeyer T (2002) Phylogenetic significance of the wing-base of the Holometabola

322	(Insecta). Zool Scripta 31:17–29.
323	Maddison DR, Maddison WP (2001) MacClade 4: Analysis of Phylogeny and Character
324	Evolution. Sinauer Inc., Sunderland.
325	Mikó I, Deans AR (2014) The second axillary in Hymenoptera. PeerJ PrePrints 2:e428v1.
326	http://dx.doi.org/10.7287/peerj.preprints.428v1
327	Misof B, Liu S, Meusemann K et al. (2014) Phylogenomics resolves the timing and pattern of
328	insect evolution. Science 346 :763–767.
329	Ogawa N, Maruyama M, Yoshizawa K (2015) Wing base morphology of Aetalionidae
330	(Hemiptera: Cicadomorpha) and its phylogenetic implications. Entomol Sci 18:262-
331	265.
332	Ouvrard D, Campbell BC, Bourgoin T, Chan KL (2000)18S rRNA secondary structure and
333	phylogenetic position of Peloridiidae (Insecta, Hemiptera). Mol Phylog Evol 16:403-
334	417.
335	Schlee D (1969) Hennig's principles of phylogenetic systematics, and 'intuitive
336	statisticophenetic taxonomy'? A reply to Dr. Coless' paper 'The phylogenetic fallacy'.
337	Syst Zool 18 :127–134.
338	Schuh RT (1979) Review: Evolutionary Trends in Heteroptera. Part II. Mouthpart-Structure
339	and Feeding Strategies. By R. H. Cobben. Syst Zool 28:653-656.
340	Sorensen JT, Campbell BC, Gill RJ, Steffen-Campbell JD (1995) Non-monophyly of
341	Auchenorrhyncha ("Homoptera"), based upon 18S rDNA phylogeny: Eco-evolutionary
342	and cladistic implications within pre-Heteropterodea Hemiptera (s.l.) and a proposal for
343	new monophyletic suborders. Pan-Pacific Entomol 71:31-60.
344	Sorenson MD, Franzosa EA (2007) TreeRot, Version 3. Boston Univ., MA.
345	Spangenberg R, Wipfler B, Friedemann K, Pohl H, Weirauch C, Hartung V, Beutel RG
346	(2013) The cephalic morphology of the Gondwanan key taxon Hackeriella
347	(Coleorrhyncha, Hemiptera). Arthropod Struct Dev 42:315-337.
348	Sweet MH (1996) Comparative external morphology of the pregenital abdomen of the
349	Hemiptera. In: Schaefer, C.W. (ed), Studies on Hemipteran Phylogeny, Entomological
350	Society of America, Maryland, pp 119–158.
351	Swofford DL (2002) PAUP*: Phylogenetic Analysis Using Parsimony (* and Other
352	Methods), Version 4. Sinauer Inc., Sunderland.
353	Urban JM, Cryan JR (2007) Evolution of the planthoppers (Insecta: Hemiptera: Fulgoroidea).

354	Mol Phylog Evol 42 :556–572.
355	Wang Y, Chen J, Jiang LY, Qiao GX (2015) Hemipteran mitochondrial genomes: Features,
356	structures and implications for phylogeny. Int J Mol Sci 16:12382-12404.
357	Wheeler WC, Schuh RT, Bang R (1993) Cladistic congruence among higher groups of
358	Heteroptera: congruence between morphological and molecular data sets. Entomol
359	Scandinavica 42 :121–137.
360	Yoshizawa K (2011) Monophyletic Polyneoptera recovered by wing base structure. Syst
361	Entomol 36 :377–394.
362	Yoshizawa K, Lienhard C (2016) Bridging the gap between chewing and sucking in the
363	hemipteroid insects: new insights from Cretaceous amber. Zootaxa 4079:229-245.
364	Yoshizawa K, Saigusa T (2001) Phylogenetic analysis of paraneopteran orders (Insecta:
365	Neoptera) based on forewing base structure, with comments on monophyly of
366	Auchenorrhyncha (Hemiptera). Syst Entomol 26:1-13.
367	Yoshizawa K, Wagatsuma M (2012) Phylogenetic relationships among superfamilies of
368	Cicadomorpha (Hemiptera: Auchenorrhyncha) inferred from the wing base structure.
369	Entomol Sci 15 :408–421.
370	
371	

Figure caption 372 373 Fig. 1. Forewing base structure of Coleorrhyncha. A. Line drawing, dorsal view. B. Image 374 taken by CLSM, dorsal view. The base of the anal vein is strongly expanded and covers 375 376 most of the membranous PMP (see also C). Note: the detachment between 3Ax and BA is an artifact caused by slide-mounting pressure (see A and C for their natural 377 relationship). C. Ventral view of wing base, showing PMP region and surrounding 378 structures. The structure in the background of the completely membranous PMP is the 379 expanded base of the anal vein (see B for comparison). 380 381 Fig. 2. Hindwing base structure of Coleorrhyncha. A. Line drawing, dorsal view. B. Image 382 taken by CLSM, dorsal view. Note: distortion in relative position of the notum and 383 axillary sclerites are an artifact caused by slide-mounting pressure (see A for their 384 natural relationship). 385 386 387 Fig. 3. The most parsimonious tree estimated from the wing base data (outgroups are omitted), with characters and their changes noted on the branched. A red square 388 indicates a non-homoplasious change, and a gray triangle indicates a homoplasious 389 change. Circled numbers are decay indices, and numbers in a square indicate 390 bootstrap/jackknife values of adjacent branches. 391 392 Fig. 4. The strict consensus of 18 equally parsimonious trees estimated from the full 393 morphological data set. See Fig. 3 for further explanations. Polytomies were treated as 394 hard polytomy for character state reconstruction. Outgroups are omitted from the figure. 395

396

- 398 Appendix: Characters and their state used for phylogenetic analyses.
- 399
- 400 **Forewing base data** (modified from Yoshizawa and Saigusa 2001)
- 401 1. Tg: (0) present; (1) absent: ci = 1, ri = 1.
- 2. Tg: (0) small; (1) enlarged, with broad extention encircling the entire margin: ci = 1, ri = 0.
- 3. Tg: (0) with small attachment to body wall; (1) with broad attachment to body wall: ci = 1,
- 404 ri = 0.
- 4. HP and BSc: (0) separate from each other; (1) united with each other: ci = 1, ri = 1.
- 5. BSc: (0) distant from 2Ax; (1) close proximity to anteroproximal corner of 2Ax; (2) fused
- with anteroproximal part of 2Ax: ci = 1, ri = 1.
- 6. BR and HP + BSc: (0) fused with each other; (1) separated from each other: ci = 1, ri = 0.
- 7. BR and 2Ax: (0) separate from each other; (1) fused: ci = 0.5, ri = 0.
- 8. 2Ax: (0) nearly flat; (1) anterior region swollen: ci = 1, ri = 1.
- 9. 2Ax: (0) not divided; (1) divided into two sclerites (2Ax-pp and -ad): ci = 1, ri = 1.
- 10. PMP: (0) located distal to 2Ax; (1) located posterodistally to 2Ax: ci = 1, ri = 1.
- 413 11. PMP: (0) nearly flat; (1) deeply concave: ci = 1, ri = 1.
- 12. PMP: (0) almost evenly sclerotized; (1) distal margin sclerotized more strongly than its
- other regions: ci = 1, ri = 1.
- 13. PMP: (0) well sclerotized; (1) reduced, often completely membranous: ci = 1, ri = 1.
- 14. DMP: (0) not divided; (1) divided into 2 sclerites: ci = 1, ri = 1.
- 15. DMP: (0) distant from 2Ax; (1) placed next to 2Ax, articulating along a convex hinge: ci
- = 1, ri = 1.
- 420 16. DMP: (0) large; (1) reduced in size: ci = 1, ri = 0.
- 17. Distal arm of 3Ax and DMP: (0) articulate with each other; (1) not articulate with each
- 422 other: ci = 1, ri = 1.
- 423 18. Anterior arm of 3Ax: (0) present; (1) absent: ci = 0.5, ri = 0.
- 424 19. 3Ax and BA: (0) separate from posterior margin of forewing base; (1) situated on
- posterior margin of forewing base: ci = 1, ri = 0.
- 426 20. BA and PMP: (0) separate from each other; (1) fused with each other: ci = 1, ri = 1.
- 427
- Full morphology data (modified from Friedmann et al. 2014)
- 1. Rupturing mechanism at the base of the antennal flagellum: (0) absent; (1) present: ci = 1,

- 430 ri = 1.
- 2. Exposure of mouthparts: (0) largely or completely exposed; (1) left mandible enclosed in a
- pouch formed by anteclypeal wall, labrum, stipes, and hypopharynx; (2) bases of
- mandibular and maxillary stylets articulate inside head with mandibular and maxillary
- 434 plates: ci = 1, ri = 1.
- 3. Right mandible: (0) present; (1) reduced: ci = 1, ri = 0.
- 4. Shape of mandibles: (0) not elongated; (1) elongated: ci = 0.5, ri = 0.92.
- 5. Cardo: (0) present; (1) strongly reduced or absent; (2) fused with stipes: ci = 1, ri = 1.
- 438 6. Lacinia: (0) absent; (1) present: ci = 1, ri = 1.
- 7. Insertion of lacinia: (0) on stipes; (1) detached from stipes: ci = 0.5, ri = 0.86.
- 8. Lacinia: (0) not elongate and stylet-like; (1) elongate and stylet-like: ci = 1, ri = 1.
- 9. Labial rostrum: (0) absent; (1) present: ci = 1, ri = 1.
- 10. Labial palps: (0) absent or strongly reduced; (1) comprising at least 2 segments: ci = 0.5,
- 443 ri = 0.86.
- 11. Cibarial water-vapour uptake apparatus: (0) absent; (1) present: ci = 0.5, ri = 0.80.
- 445 12. Jugal "bar": (0) absent; (1) present: ci = 1, ri = 1.
- 13. Abdominal ganglia: (0) more than two separate ganglia; (1) two separate ganglia; (2) one
- single ganglionic mass: ci = 1, ri = 1.
- 14. Eyes of immature stages: (0) persist; (1) disintegrate or pulled back proximally into
- 449 cerebrum: ci = 1, ri = 1.
- 450 15. External wing buds: (0) present; (1) absent: ci = 1, ri = 1.
- 451 16. Pupal stage: (0) absent; (2) present: ci = 0.5, ri = 0.5.
- 17. Appearance of compound eyes: (0) before ultimate immature stage; (1) in ultimate
- immature stage: ci = 1, ri = 1.
- 18. Ocelli of immature stages: (0) present; (1) absent: ci = 1, ri = 1.
- 455 19. Cerci of immature stages: (0) present; (1) absent: ci = 1, ri = 1.
- 456 20. Tegulae of the forewing: (0) present; (1) absent: ci = 1, ri = 1.
- 21. Size and shape of tegulae: (0) small; (1) enlarged, with broad extension encircling the
- 458 entire margin: ci = 1, ri = 1.
- 459 22. Attachment of tegulae to body wall: (0) narrow; (1) broad: ci = 0.5, ri = 0.
- 460 23. HP and BSc: (0) separated from each other; (1) connected with each other: ci = 1, ri = 0.
- 24. BSc: (0) distant from 2Ax; (1) closely adjacent with the anteroproximal corner of 2Ax;

- 462 (2) fused with anteroproximal part of 2Ax: ci = 0.67, ri = 0.67.
- 25. BR and HP + BSc: (0) fused with each other; (1) separated from each other: ci = 1, ri = 0.
- 26. BR and 2Ax: (0) separated from each other; (1) fused: ci = 0.5, ri = 0.
- 465 27. 2Ax: (0) nearly flat; (1) anterior region inflated: ci = 1, ri = 1.
- 466 28. 2Ax: (0) not divided; (1) not divided: ci = 1, ri = 1.
- 29. Position of PMP: (0) distad 2Ax; (1) posterodistad 2Ax: ci = 1, ri = 0.
- 468 30. PMP: (0) nearly flat; (1) deeply concave: ci = 1, ri = 0.
- 31. PMP: (0) almost evenly sclerotized; (1) distal margin sclerotized more strongly than its
- other regions: ci = 1, ri = 0.
- 32. PMP: (0) well sclerotized; (1) reduced, often completely membranous: ci = 0.5, ri = 0.86.
- 33. DMP: (0) not divided; (1) divided into 2 sclerites: ci = 1, ri = 0.
- 34. DMP: (0) distant from 2Ax; (1) placed next to 2Ax, articulating along a convex hinge: ci
- = 1, ri = 1.
- 35. DMP: (0) large; (1) reduced in size: ci = 1, ri = 0.
- 36. Distal arm of 3Ax and DMP: (0) articulating with each other; (1) not articulating with
- each other: ci = 1, ri = 0.
- 478 37. Anterior arm of 3Ax: (0) present; (1) absent: ci = 0.5, ri = 0.
- 38. 3Ax and BA: (0) separate from posterior margin of forewing base; (1) attached to
- posterior margin of forewing base: ci = 1, ri = 0.
- 39. BA and PMP: (0) separate from each other; (1) fused with each other: ci = 1, ri = 0.
- 482 40. Lateral hypopharyngeal arm (0) present; (1) absent: ci = 0.5, ri = 0.88.
- 41. Ovarioles: (0) not polytrophic; (1) polytrophic; (2) telotrophic; (3) panoistic: ci = 0.75, ri
- 484 = 0.92.
- 485 42. Maxillary palps: (0) present, with four segments or more; (1) absent or reduced number of
- 486 segments: ci = 0.33, ri = 0.8.
- 43. Abdominal sternite 1: (0) present; (1) absent: ci = 0.5, ri = 0.8.
- 488 44. Number of axonemes in spermatozoans: (0) zero; (1) one; (2) two; (3) three: ci = 0.75, ri
- 489 = 0.83.
- 490 45. Gonangulum: (0) not fused with tergum IX; (1) fused with tergum IX: ci = 0.5, ri = 0.8.
- 491 46. Pretentorium: (0) absent or if present not connecting internal extremities of mandibular
- lever and corpotentorium; (1) unites internal extremities of mandibular lever and
- 493 corpotentorium: ci = 1, ri = 1.

- 494 47. Lacinial gland: (0) absent; (1) present: ci = 0.5, ri = 0.
- 48. Male genitalia: (0) symmetrical, or if asymmetrical, asymmetry not involving pregenital
- segments; (1) asymmetrical, this asymmetry often involving pregenital segments: ci =
- 497 0.5, ri = 0.
- 498 49. Accessory salivary glands generally: (0) not tubular; (1) of the tubular type: ci = 1, ri = 0.
- 50. Number of eye trichobothria of first instars: (0) one or absent; (1) two: ci = 1, ri = 0.
- 51. Number of ommatidia in first-instar larvae: (0) 4-5; (1) more than five: ci = 0.5, ri = 0.
- 501 52. Number of tarsomeres in first-instar larvae: (0) one; (1) two: ci = 1, ri = 1.
- 53. Forewings: (0) completely uniform or if differentiated, not forming a distinct
- corium-clavus and membrane; (1) forewing divided into a distinct corium-clavus and
- 504 membrane: ci = 0.5, ri = 0.
- 505 54. Cephalic trichobothria: (0) absent in adults; (1) present in adults: ci = 1, ri = 1.
- 506 55. Metathoracic scent gland system: (0) absent; (1) present: ci = 1, ri = 1.
- 56. Labium: (0) not tubular; (1) tubular labium with three segments; (2) tubular labium with
- four segments: ci = 1, ri = 1.
- 509 57. Insertion of tubular labium: (0) posteriorly on the head, (1) anteriorly on the head: ci = 1,
- ri = 1.
- 58. Dorsal abdominal glands in immature stages: (0) absent; (1) present: ci = 1, ri = 1.
- 59. Number of antennal flagellomeres: (0) more than 4, (1) 4 or less: ci = 0.33, ri = 0.75.
- 60. Articulations between the mesomere, anterodorsal extension of ventral plate and posterior
- end of basal plate: (0) absent; (1) present: ci = 0.5, ri = 0.5.
- 61. Length of basal apodeme of the phallic organ: (0) short; (1) long, longer than basal plate:
- 516 ci = 1, ri = 0.
- 62. Third posterodorsal corner of basal plate: (0) not extended; (1) extended posteriorly: ci =
- 518 1, ri = 0.
- 63. Basal apodeme of the phallic organ: (0) present; (1) absent: ci = 1, ri = 1.
- 64. Width of basal apodeme: (0) narrow; (1) as broad as or broader than basal plate: ci = 1, ri
- 521 = 1.
- 522 65. Ventral plates 1: (0) separated; (1) partly fused anteriorly: ci = 1, ri = 1.
- 66. Ventral plates 2: (0) separated or partly fused; (1) completely fused: ci = 1, ri = 0.
- 67. Mesomere of the aedeagus: (0) rounded posteriorly; (1) pointed posteriorly: ci = 0.5, ri =
- 525 0.5.

- 68. Posteromedian part of basal plate: (0) membranous; (1) sclerotized: ci = 1, ri = 1.
- 69. Anterior end of mesomere: (0) articulated with basal plate; (1) articulated with paramere:
- 528 ci = 1, ri = 0.
- 70. Paired ocelli in nymphs or larvae: (0) absent; (1) present: ci = 1, ri = 1.
- 71.Intrinsic antennal muscles (Mm. scapopedicellares) in immature stages: (0) absent; (1)
- 531 present: ci = 1, ri = 1.
- 72. Ventral metasternal process: (0) absent; (1) present: ci = 1, ri = 1.
- 73. Sensory plate organs of pedicel: (0) absent; (1) present: ci = 1, ri = 1.
- 534 74. Evan's organ: (0) absent, (1) present: ci = 0.5, ri = 0.86.
- 75. Ductus ejaculatorius: (0) normal; (1) modified as a sperm pump: ci = 1, ri = 1.
- 76. Proximal abdomen pediculate by reduction of the 1st and 2nd segment: (0) absent; (1)
- 537 present: ci = 1, ri = 1.
- 538 77. Hind coxae: (0) normally developed; (1) broad, closely adjacent: ci = 1, ri = 1.
- 78. Proboscis: (0) absent; (1) shifted posteriorly between bases of procoxae; (2) not shifted
- posteriorly between bases of procoxae: ci = 0.67, ri = 0.94.
- 79. Posterior parts of the head capsule: (0) sclerotized; (1) membraneous: ci = 1, ri = 1.
- 80. Connective tissue occluding occipital foramen: (0) absent; (1) present: ci = 0.5, ri = 0.
- 81. Ovipositor simplified: (0) absent; (1) present: ci = 0.5, ri = 0.5.
- 82. Spiracular glands: (0) absent; (1) present: ci = 0.5, ri = 0.
- 83. Extension of the occipital apodeme reaching into the thorax: (0) absent; (1) present: ci =
- 546 0.5, ri = 0.
- 84. Pronotum and procoxae: (0) not fused; (1) fused: ci = 1, ri = 1.
- 85. Position of anterior tentorial pits: (0) frontal side of head; (1) absent; (2) shifted dorsally:
- 549 ci = 1, ri = 1.
- 86. Fusion of head and thorax: (0) absent; (1) present: ci = 1, ri = 1.
- 87. Body and head: (0) not flattened; (1) dorsoventrally flattened: ci = 0.25, ri = 0.67.
- 88. Hind femora: (0) not enlarged; (1) enlarged: ci = 0.33, ri = 0.6.
- 89. Meso- and metanotum: (0) not fused; (1) fused: ci = 1, ri = 1.
- 554 90. Compound eyes: (0) not reduced; (1) only 2 ommatidia or less: ci = 0.5, ri = 0.86.
- 91. Labial palp: (0) present; (1) absent: ci = 0.5, ri = 0.88.
- 556 92. Complex tymbal acoustic system: absent (0); present (1): ci = 1, ri = 1.
- 93. Aristate antennal flagellum: (0) absent; (1) present: ci = 0.5, ri = 0.86.

- 94. Malpighian tubules: (0) more than six; (1) six; (2) four or less: ci = 1, ri = 1.
- 95. Labrum: (0) not narrowed; (1) narrowed: ci = 1, ri = 1.
- 96. Mandibular and lacinial stylets: (1) unicondylar; (0) dicondylar: ci = 1, ri = 1.
- 97. Pedunculate eggs (with stalk): (0) absent; (1) present: ci = 1, ri = 1.
- 98. Gut with filter chamber containing Malpighian tubules: (0) absent; (1) present: ci = 1, ri =
- 563 1.
- 99. Coronal (= median epicranial) suture: (0) absent; (1) present: ci = 0.33, ri = 0.33.
- 100. Parempodia on unguitractor plate: (0) absent; (1) elongate and setiform, inserted in an
- size $c_i = 0.5$, $c_i = 0.5$.
- 101. Number of tarsal segments: (0) one; (1) two; (2) three; (3) more than three. State 3 is
- adopted for Mydiognathus: ci = 0.33, ri = 0.65.
- 569 102. Arolium: (0) absent; (1) present; (2) eversible; (3) bilobed: ci = 0.33, ri = 0.45.
- 570 103. Sticky terminal lip of arolium: (0) absent; (1) present: ci = 0.5, ri = 0.67.
- 571 104. Pulvilli: (0) absent; (1) present: ci = 0.33, ri = 0.6.
- 572 105. Euplantulae: (0) absent, (1) present: ci = 0.33, ri = 0.33.
- 573 106. Number of claws: (0) one; (1) two; (2) reduced into spoon-shaped plates; (3) main claw
- plus accessory claw: ci = 0.75, ri = 0.67.
- 575 107. Claw teeth: (0) absent; (1) present: ci = 0.33, ri = 0.33.
- 576 108. Protuberance with microtrichia on distolateral side of the pretarsus: (0) absent; (1)
- 577 present: ci = 1, ri = 0.
- 578 109. Sensorial setae on mesal side of arolium: (0) absent; (1) present: ci = 1, ri = 1.
- 579 110. Adhesive claw setae: (0) absent; (1) present: ci = 1, ri = 0.
- 580 111. Eversible structure between tibia and tarsus: (0) absent; (1) present: ci = 0, ri = 0.
- 581 112. Tibial thumb-like process: (0) absent; (1) present: ci = 1, ri = 0.
- 582 113. Empodial paronychium: (0) absent; (1) present: ci = 1, ri = 0.
- 583 114. Tarsal apophysis on the ventral side of the tarsus: (0) absent; (1) present: ci = 1, ri = 0.
- 584 115. Two dorsal capitate setae: (0) absent; (1) present: ci = 1, ri = 0.
- 585 116. Flag-like sensilla on the 1st tarsal segment: (0) absent; (1) present: ci = 1, ri = 0.
- 586 117. Fingerlike process below claw: (0) absent; (1) present: ci = 1, ri = 0.
- 587 118. Ventral brush: (0) absent; (1) present: ci = 0.5, ri = 0.









