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3 Co-phylogeography and morphological evolution of sika deer lice

4 (*Damalinia sika*) with their hosts (*Cervus nippon*)

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## 22 SUMMARY

23 Lice are obligate parasites of mammals and birds and have become an important  
24 model for studies of host-parasite coevolution and cophylogenetics. Population  
25 genetic and phylogeographic studies represent an important bridge between  
26 microevolution and cophylogenetic patterns. We examine co-phylogeographic  
27 patterns in sika deer and their parasitic lice. Co-phylogeographic patterns in deer  
28 and lice were evaluated using homologous regions of mitochondrial COI sequences.  
29 The phylogeographic breaks recovered for deer populations matched those of  
30 previous studies. Comparisons of the phylogeographic tree topology for deer lice  
31 with that of their hosts revealed a significant level of congruence. However,  
32 comparisons of genetic distances between deer and lice suggested one of the  
33 estimated co-divergence events is more likely a recent host switch. Taking into  
34 account genetic divergence, there is not strong evidence for complete  
35 phylogeographic co-divergence between deer and their parasitic lice. However,  
36 mitochondrial phylogenies only track genetic structure of female lineages, and the  
37 incongruence between deer and louse phylogeography may be explained by louse  
38 migration mediated by male deer. Morphological analysis of head shape variation  
39 based on an elliptic Fourier descriptor showed that overall morphological variation  
40 contained phylogenetic signal, suggesting that in general morphology of these lice  
41 evolves congruent to population history.

42

43 Key words: co-phylogeography, population structure, microevolution, morphology,  
44 parasitic louse, mammal

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47 INTRODUCTION

48 Parasitic lice (Insecta: Psocodea: "Phthiraptera") are obligate parasites of  
49 mammals and birds and spend their entire lifecycle on the hosts. Because of this  
50 permanent association with their hosts, lice have been used as a model system for  
51 studies of host-parasite codivergence (Page, 2002). Several studies comparing  
52 louse and host phylogenies have provided evidence for codivergence between lice  
53 and their hosts (e.g., Hafner *et al.* 1994; Page *et al.* 1998; Johnson *et al.* 2002*b*;  
54 Light and Hafner, 2008; Smith *et al.* 2008). In contrast, several cases in which  
55 substantial incongruence between host and parasite trees exists have been  
56 identified (Johnson *et al.* 2002*a,b*; Weckstein, 2004; Banks *et al.* 2006). All of  
57 these studies have focused on species level phylogenies, and there are only a few  
58 studies focusing on codivergence within species, among populations (Whiteman *et*  
59 *al.* 2007; ˇStefka *et al.* 2011). Thus, it is still generally unclear whether  
60 codivergence at the species level is driven by population level codivergence that  
61 precedes speciation. Population genetic and phylogeographic studies will be an  
62 important bridge between microevolution and cophylogenetic patterns (Johnson *et*  
63 *al.* 2003*a*).

64 To understand the causes of congruence and incongruence between host and  
65 parasite phylogenies, careful consideration of host and parasite behaviors, effects of  
66 other members of the parasite community (e.g. phoresis by hippoboscids flies), and  
67 geographical information are needed (Johnson *et al.* 2002*a,b*). As long as these  
68 factors are accounted for, parasite phylogeny could also provide insights regarding  
69 the host's phylogeny (Hopkins, 1942; Reed *et al.* 2004; Johnson *et al.* 2006;  
70 Whiteman *et al.* 2007). In particular, the molecular substitution rate for lice is  
71 generally much faster than that of their hosts (Hafner *et al.* 1994; Huelsenbeck *et*  
72 *al.* 1997; Page, 1996; Page *et al.* 1998; Johnson *et al.* 2003*b*), and substitutional  
73 differences may accumulate between diverging louse populations at shallow  
74 timescales even though their hosts have not yet accumulated any mutational  
75 differences. Therefore, population genetic and phylogeographic studies of parasitic  
76 lice also have the potential to uncover hidden population structure in their hosts.

77 In addition to studies of codivergence, lice have also been used as an  
78 important system in studying morphological evolution of coevolving systems. The  
79 evolution of morphological traits can be affected by many factors, such as  
80 phylogenetic constraints, gene flow, genetic drift, sexual selection, and, most  
81 importantly, selection pressure from the environment. Generally, it is very difficult  
82 to separate these factors clearly. Parasitic lice spend their entire life cycle on the  
83 body of the host and thus the environmental context for their morphological  
84 evolution can be more easily identified than that for free-living organisms. For this  
85 reason, lice are good models for studying the impact of the host on morphological  
86 evolution of these parasites. Previous studies have shown interesting  
87 coevolutionary patterns between lice and their hosts. For example, Reed *et al.*  
88 (2000) clearly showed that the head groove width of gopher chewing lice is strongly  
89 correlated with the hair diameter of the host. Mammal lice hang onto the host hair  
90 using the head groove, so that a mismatch between groove size and hair diameter  
91 likely increases the risk to the louse of falling off or being groomed off the host.  
92 Johnson *et al.* (2005) also found that the body width of wing lice from pigeons and  
93 doves is strongly correlated with host body size. Host body size dictates the width  
94 of the feather interbarb space, in which wing lice insert themselves to escape host  
95 preening defenses. As in the case of the gopher lice, a mismatch between the body  
96 width of the louse and the feather interbarb space increases the risk of being  
97 preened off the host (Clayton *et al.* 2003; Bush and Clayton, 2006).

98 Most previous studies of louse morphological evolution have focused only on  
99 functionally significant characters, such as louse body width vs feather interbarb  
100 space or louse head groove size vs hair diameter. However, the functional  
101 significance (if any) of many taxonomically and phylogenetically useful morphological  
102 characters in lice is not clear. In addition, recent progress on the higher level  
103 molecular phylogeny of parasitic lice has revealed that morphological convergence  
104 can be very frequent for characters used in louse taxonomy and phylogeny  
105 reconstruction (Johnson *et al.* 2004, 2012; Smith *et al.* 2004; Yoshizawa and  
106 Johnson, 2006, 2010). For lice, little is known about how these taxonomically and

107 phylogenetically useful morphological characters vary across populations and how  
 108 these characters evolve over microevolutionary timescales. Such knowledge would  
 109 be useful to understand the evolution of morphology over macroevolutionary time.  
 110 Quantitative comparisons of morphometric data and phylogeny are an effective way  
 111 to study these characters (Klingenberg and Gidaszewski, 2010), but no such study  
 112 has been done for lice to date.

113 Here, we undertake a co-phylogeographic study of sika deer and their  
 114 parasitic lice. The sika deer (*Cervus nippon*) is distributed in east Asia and  
 115 throughout the Japanese Archipelago. Japanese populations are subdivided into six  
 116 subspecies and are highly variable in body size (from 30 to 120 kg in male's body  
 117 mass: Ohtaishi, 1986). Such a large variation in body size within a species of  
 118 mammal is exceptionally rare (Ohdachi *et al.* 2009). However, morphology-based  
 119 subspecific subdivisions within sika deer are not supported by mitochondrial  
 120 phylogeny, and two distinct mitochondrial lineages have been identified for  
 121 Japanese sika deer. Sika deer host a parthenogenetic chewing louse species,  
 122 *Damalinia sika*, making this a potentially ideal system to evaluate patterns of co-  
 123 phylogeography and morphological evolution. As mentioned above, morphological  
 124 evolution can be affected by many factors, including gene flow via interbreeding. *D.*  
 125 *sika* is an obligately parthenogenetic louse, and thus gene flow via interbreeding  
 126 among populations can be excluded as a factor affecting morphological evolution,  
 127 making this species useful for distinguishing other factors impacting morphological  
 128 evolution of this species.

129 In this study, we estimated the mitochondrial population level phylogenies of  
 130 sika deer and their lice, based on sequences of the COI gene. We investigated co-  
 131 phylogeographic patterns by comparing tree topologies and genetic divergence  
 132 between hosts and parasites. We also discuss novel insights regarding deer  
 133 population structure based on phylogeographic patterns observed in their lice. To  
 134 examine patterns of louse morphological evolution, we analyzed head shape using  
 135 morphometric analyses. We compared the louse population tree from COI  
 136 sequences with the results of cluster analysis of morphometric data.

137

## 138 MATERIALS AND METHODS

139 *Samples*

140 Deer and deer louse samples were collected throughout the Japanese  
141 archipelago (Fig. 1). Most louse samples were collected by the authors and fixed in  
142 99% ethanol immediately. Hairs of deer were also sampled with lice and were stored  
143 in 99% ethanol. Some deer samples were collected by local hunters, and frozen  
144 skins were sent to the authors. Lice were collected from the skins and fixed with  
145 99% ethanol. Numbers of deer and lice samples examined are listed in Table 1. In  
146 most cases, samples of lice came from the individual deer included in this study.  
147 However, the Taiwanese population of louse was represented from a host individual  
148 that was unavailable for sequencing. Sequences from the host in this case (*C. n.*  
149 *taionus*) was represented by sequences obtained from GenBank. In addition,  
150 sequences from the Far East Russian population of sika deer were obtained from  
151 GenBank and included in the phylogeographic analysis of deer, even though louse  
152 samples were not available for analysis. This was done to stabilize the tree search  
153 by breaking the long branch leading to Taiwanese sika deer (Table 1). *Cervus*  
154 *elaphus* and *Damalinia forficula* were used as outgroups for the host and parasite  
155 trees, respectively. These species are not associated, but are simply used for  
156 rooting the ingroup topologies because they are among the closest relatives of *C.*  
157 *nippon* and *D. sika*.

158 Sequences of a portion of the mitochondrial COI gene were obtained for both  
159 deer (838 bp) and lice (776 bp). Though the length of these sequences were slightly  
160 different, the sequence for the deer completely overlapped that for the lice, making  
161 these regions homologous. Therefore, the sequence data can be utilized for the  
162 comparison of host-parasite substitution rates (see below). The mitochondrial  
163 control region is known to evolve rapidly and thus is commonly used for  
164 phylogeography of mammals (Nagata *et al.* 1999). However, because mitochondrial  
165 genome of deer lice is composed of minicircles (Cameron *et al.* 2011), homology of  
166 the control regions between deer and deer lice cannot be warranted. No nuclear

167 markers for phylogeographic studies are known for deer (Barbosa and Carranza,  
168 2010). Therefore, to compare homologous gene regions between hosts and  
169 parasites, COI is the best option to date. In addition, because sika deer lice are  
170 parthenogenetic, mitochondrial DNA is one of the best markers for tracking the  
171 history of these parthenogenetic lines. Procedures for total DNA extraction, PCR,  
172 and sequencing followed Yoshizawa and Johnson (2010), but primers were newly  
173 designed here (supplementary data). Alignment was done by eye because of the  
174 lack of indels in this protein coding gene. The aligned matrix in nexus format and  
175 csv file for statistical analyses (see below) are available as Supplementary Data.  
176 Sequence data also are deposited at GenBank (Table 1). Uncorrected and  
177 likelihood-based genetic distances were calculated using PAUP\* 4b10 (Swofford,  
178 2002).

179

#### 180 *Phylogeographic Analyses*

181 Phylogeographic analyses were performed with the parsimony (PAUP\*),  
182 likelihood (PAUP\* and PhyML 3.0: Guindon *et al.* 2010), and Bayesian (MrBayes  
183 3.12: Ronquist and Huelsenbeck, 2003) criteria. Best fit models for the ML and  
184 Bayesian analyses were calculated by hLRT using Modeltest (Posada and Crandall,  
185 1998) and MrModeltest (Nylander, 2004), respectively (Supplementary Data). For  
186 ML analyses, using PAUP\* a heuristic search with TBR branch swapping was  
187 implemented for each data set, using a NJ starting tree. For MP analysis, 100  
188 random addition replicates using TBR heuristic branch swapping were performed.  
189 PAUP\* was also used for MP bootstrapping, with 1000 replicates. PhyML was used  
190 to conduct likelihood-based 1,000 bootstrap replicates, using NNI branch  
191 swapping, each with Bio-NJ as a starting tree. For Bayesian analyses, we performed  
192 2 runs, each with 4 chains for 10,000,000 generations, and trees were sampled  
193 every 1,000 generations. For each run, the first 5,000 trees were excluded as a  
194 burnin and the 50% majority consensus tree of the remaining trees was calculated to  
195 estimate posterior probabilities. We also estimated ParsimonySplits networks as  
196 implemented in SplitsTree4 (Huson and Bryant, 2006) using default settings. The

197 outgroups were excluded from the splits analyses.

198

#### 199 *Codivergence Analyses*

200 Codivergence analysis was performed using the Jungle algorithm (Charleston,  
201 1998) in TreeMap 2.02b (Page and Charleston, 1998). Samples with identical  
202 haplotypes were trimmed to a single terminal taxon. All non-codivergence events  
203 were given equal costs of 1 and codivergence was given a cost of 0. As discussed  
204 below, a number of non-codivergence events were identified from the Jungle  
205 analysis. To test whether there was significant evidence for a lack of strict  
206 codivergence, some alternative genealogical topologies were tested using the  
207 approximately unbiased test (AU test: Shimodaira, 2002) using CONSEL  
208 (Shimodaira and Hasegawa, 2001). The partition homogeneity test (Farris *et al.*,  
209 1994, 1995) was also performed using PAUP\* to evaluate whether host and parasite  
210 data sets represented significantly different topologies (Johnson *et al.* 2001). To  
211 perform the test, a data matrix in which hosts and parasites were matched in 1:1  
212 associations (as described by Johnson *et al.* 2001) was prepared by excluding all the  
213 samples lacking a host-parasite association (i.e., outgroups and sika deer from  
214 Ussuri) and also eliminating the samples for which both the hosts and parasites from  
215 an identical locality show exactly identical haplotypes. Using this 1:1 data set,  
216 1,000 replicates of the partition homogeneity tests were performed each with TBR  
217 branch swapping. This 1:1 association data set was also used to compare the  
218 genetic distance between deer and lice. PAUP\* was used to estimate the genetic  
219 distances, and uncorrected pairwise distances of total data, uncorrected pairwise  
220 distances of four-fold degenerate sites, and likelihood-based distances using the  
221 models estimated above were calculated. MEGA5 (Tamura *et al.* 2011) was used to  
222 identify four-fold degenerate sites.

223

#### 224 *Analyses of Louse Morphology*

225 For parasites, host imposed selection may be more important than the  
226 parasite population or phylogenetic history for shaping the patterns of



227 morphological evolution. To evaluate the correspondence between louse  
228 morphology and population structure of both deer and lice, we analyzed the shape  
229 of the louse head. We used elliptic Fourier analysis (Kuhl and Giardiana, 1982) as  
230 implemented in the software package SHAPE (Iwata and Ukai, 2002). Morphological  
231 terminology follows Lyal (1985).

232 For morphological analysis, we examined 138 louse specimens from 15  
233 localities, and each locality was treated as a terminal. The morphological samples  
234 did not include those used for DNA extraction when Proteinase K during the  
235 extraction caused breakage of the head capsule. Specimens that were used only for  
236 morphological analyses were cleared using 10% KOH solution for one night at room  
237 temperature. All specimens were rinsed with 80% ethanol (3 times), dehydrated with  
238 99.5% ethanol (2 times), and slide mounted with Euparal.

239 The head capsule was photographed through the microscope using a digital  
240 camera. Using the digital image, the head outline was traced and scanned. Because  
241 head shape of *D. sika* is close to a circle, the initial alignment of head shape had to  
242 be re-orientated. To do this, the right margin of the outline was oriented on the  
243 right margin of ventral carina. Following Smith (2000), 20 harmonics were  
244 calculated from the digitized outline, while excluding effects of size. The resulting  
245 80 coefficients were subjected to principal component analysis using SHAPE. The  
246 13 principal component scores (PC 1–13) that represented over 95% of the  
247 variation were used for canonical discriminant analysis. We then calculated  
248 Mahalanobis distances between each group of specimens from the same locality.  
249 Then, using the Mahalanobis distances among locality groups, we performed cluster  
250 analysis using the group average method. Discriminant and cluster analyses were  
251 performed using JMP 9.0 (SAS, 2011).

252

253 RESULTS

254 *Phylogeography of Deer and Deer Lice*

255 ML analysis of each data set recovered a single most likely tree (Figs 2–3).  
256 MP analyses recovered 18 (for deer) and 11 (for louse) trees of equal length, and

one of these was the same as the tree estimated from the ML analyses.

ParsimonySplits networks are provided as Supplementary Figs 1–2.

Three major lineages (North, South, and Continental + Taiwanese Clades) were recovered for sika deer (Fig. 2, Suppl. Fig. 1). Support values for all clades were generally strong (>77% BS for all clades and 98% PP for South Clade), but only low Bayesian support values were obtained for North (98–100% BS vs 82% PP) and Continental + Taiwanese Clades (77–83 BS vs 81% PP). Two Japanese lineages (Northern and Southern lineages) composed a clade, and the Continental + Taiwanese samples formed the other clade. However, resolution of the basal relationships among these lineages was weakly supported (59% BS and 56% PP). Low resolution of the basal divergences was also evident from the splits network (Suppl. Fig. 1). Analysis of the smaller data set (1:1 data, see Materials and Methods) placed Taiwanese sika deer as the sister of the Southern clade, making Japanese sika deer paraphyletic (Supplementary Data). The Southern Clade included all samples from Kyushu and its adjacent islands. The Northern Clade consisted of the samples from Hokkaido and Honshu.

The analyses of the louse data set recovered two distinct lineages (Clades I and II: Fig. 3 and Suppl. Fig. 2). Support values for both clades were very strong. Clade I was subdivided into two clades (Ia and Ib). Clade Ia was well supported and included samples from Hokkaido, northern Honshu and Fukuoka. The basal branch of Clade Ib was extremely short and weakly supported. This clade was composed of samples from Nikko, southern Honshu, Tsushima and Kagoshima. In the split network, samples of Tsushima were placed closer to Clade Ia than the other samples of Clade Ib, although network connecting them was extremely short (Suppl. Fig. 2). Clade II was composed of the samples from southern Kyushu and adjacent islands, and the sample from Taiwan.

#### *Codivergence Analysis*

Initial Jungle analysis using TreeMap 2 recovered four equally optimal results with four codivergence events and 13 non-codivergence events (Figs 4–5).

Standard randomization test showed that the number of codivergence events was significantly higher than that expected by chance in 1000 randomizations of parasite tree ( $P=0.005$ ). However, most cophylogenetic analyses do not accommodate parasites that occur on multiple host terminals, such as a single species of parasites on multiple host species or parasite populations found on multiple host populations (Johnson *et al.* 2002a). Analytically these are often *post hoc* considered to be recent host switching events. Because of these difficulties in dealing with widespread parasites, the results from the TreeMap analysis could not account for two shallow host switching events of lice from deer in Fukuoka to North1 and to Toya populations (asterisks in Fig. 5). There still was significant congruence between host and parasite trees ( $P=0.034$ ) even if these two additional non-codivergence events were allowed.

Because louse phylogeography did not closely correspond to host deer divergence patterns, we also examined these topological incongruences statistically. When the distinction between the South and North populations in the deer was forced as a constraint on the louse tree (excluding the Taiwanese deer louse, which was kept within the Southern Clade, together with Kyushu deer louse, as in the original louse phylogeny), the resulting tree was significantly worse than the unconstrained tree using the AU test ( $P<0.001$ ). In contrast, when only the basal divergence between the Taiwanese and Japanese populations as estimated from the deer phylogeny was constrained in the louse tree, this constraint tree was not significantly worse than the original using the AU test ( $P=0.217$ ). Similarly for the deer, when the two major clades identified by the phylogenetic analyses of the louse was constrained in the tree for sika deer (excluding the Taiwanese sika deer which was kept separate from Japanese population as in the original deer phylogeny), the constrained tree was significantly worse using the AU test ( $P=0.027$ ). Likewise, the partition homogeneity test recovered significant heterogeneity between the host-parasite COI sequences ( $P=0.001$ ).

315

316 *Comparisons of Genetic Distances*

Fig. 6 shows plots of the uncorrected p-distances between pairs of lice and those from their deer hosts. These plots contain roughly five clusters of points (Groups A–E). The divergence events of deer and lice corresponding to the plot groups are indicated by shade in the figure, and the plots corresponding to the codivergence events are indicated by circled numbers (Figs 2–3, 5–6). The uncorrected p-distances of four-fold degenerate sites and likelihood-based distances also showed identical trends (not shown). When points in groups B and C were excluded (because they clearly represent some incongruence between louse and deer population histories) and correlation between deer and louse genetic distances were calculated, the substitution rate of the COI gene for lice was estimated to be about 4.8 times faster than that of deer (Fig. 6). This estimate also accords with the fact that the highest divergences among lice are around 12%, while the highest among deer are only 2.5%. Although plot group E corresponds to the co-divergence event between Kumamoto and Yakushima + Kuchinoerabu populations (Fig. 5), these points are also outliers of the estimated slope (Fig. 6).

### *Morphological Variation of Lice*

Except for the populations from Kinkazan and Nara, all clusters identified by morphological analysis corresponded to the lineages recovered from the louse molecular phylogenetic analysis (Figs 7–8). Note that molecular phylogenetic analysis divided the Nikko population into two distinct lineages (Fig. 3), but the majority of these specimens were placed in the Clade Ia, which corresponded to the cluster identified by morphological analysis. In contrast, except for two populations collected from the same subspecies of deer (i.e., *C. n. yakushimae*: note that lice from these two populations are also genetically closely related: Fig. 3), the clusters identified from louse head morphology did not show any grouping corresponding to deer subspecies or geography.

PC1 explained variation in the shape of conus, depth of the antennal groove, head width, and width of the pulvinus (Fig. 7: proportion = 32.6%). PC2 also explained variation in the protrusion of the antennal groove margin, protrusion of

the temple, shape of the conus, and width of the pulvinus (proportion = 26.9%). PC3 explained variation in the antennal groove (proportion = 10.1%). PC4 explained clear variation in the depth of the pulvinus, and explained some variation in the depth of the antennal groove (proportion = 9.1%). The proportion of morphological variation explained by PC5–13 was less than 5% (not shown). However, PC5 (proportion = 4.7%) still contained detectable variation in the depth of the antennal groove, and PC6 (proportion = 2.7%) explained clear variation in the length of the conus. PC7–13 explained very little of the overall variation.

## DISCUSSION

### *Codivergence between Sika Deer and Lice*

Comparing deer and louse trees using Jungle analyses recovered significant patterns of topological congruence. However, these analyses only account for the branching pattern and do not take into account the relative timing of branching events. Using information from the genetic divergence plots, three of four reconstructed codivergent events are also concordant with the relative timing of branching events (1, 3 and 4 in Fig. 6), providing reciprocal support for the results of both types of analyses. However, plot Group E, which would correspond to a codivergence between Kumamoto and Yakushima + Kuchinoerabu population (2 in Fig. 6), is apparently an outlier for genetic distance as compared to the other codivergence events. Therefore, even though the branching patterns of these populations might indicate codivergence, based on genetic distances this is more likely a recent host switch. When one of the codivergence events was excluded (i.e., number of codivergence events was set to three and one more host switching event was allowed), the tree randomization test showed that the number of codivergence events was not significantly higher than that expected by chance ( $P=0.19$ ). Significant mismatches between the deer and louse trees are also evident from the plots of the genetic distances, the AU test, and the partition homogeneity test.

In contrast, the AU test did not reject the possibility of basal diversification

of the Taiwanese deer louse from the Japanese population. Alternatively, when the smaller data set was analyzed, the Taiwanese sika deer was placed to the sister of the Southern Clade, agreeing with the louse tree as far as the placement of Taiwanese population is concerned. However, there is a significant discordance between pairwise genetic distance comparisons for these deer and louse populations (plot group C in Fig. 6). Therefore, by considering both topology and genetic distances, the close relationship between louse populations of Taiwan and southern Kyushu is unlikely to be a result of their codivergence and, as estimated from Jungle analyses, this should also be interpreted as a result of a recent host switch.

The louse populations in Kyushu are highly structured genetically, even within Clade I, whereas those of Hokkaido and Honshu show much less genetic variability (Fig. 3). Jungle analysis suggests that the louse populations in Hokkaido and Honshu originated via a recent host switch from deer in Kyushu. The deer populations in Honshu and Hokkaido show some genetic structure (Fig. 2), but the genetic variation of lice in Honshu and Hokkaido is apparently less structured and less variable (Fig. 3), even though the substitution rates of lice are much faster than those for deer (Fig. 6). The lack of genetic variability and structure in these louse populations also supports the hypothesis, from Jungles analysis, that they originated from a recent host switch, which may have been associated with a genetic bottleneck.

The evidence for multiple host switching events suggests that deer lice migrate among deer populations quite readily and independent of deer population structure. Mismatches between host and louse phylogenies at species level are quite frequent for bird lice (Johnson *et al.* 2002*a,b*; Weckstein, 2004; Banks *et al.* 2006). In contrast, all previous studies of mammal lice have identified highly significant codivergence patterns between mammal lice and their hosts at the species (Hafner *et al.* 1994; Light and Hafner, 2007; Light *et al.* 2008; Smith *et al.* 2008) or population levels (Demastes *et al.* 2012). Although trends similar to other mammal lice were expected in these deer lice, it appears that different dynamics may operate for the deer–lice system, and these are explored below.

407

408 *Phylogeography of Deer Lice*

409       The most basal divergence within deer lice was estimated to correspond to  
 410 the divergence between continental and Japanese deer lineages (Figs 2–3). Clade II  
 411 appears to have its ancestor in Taiwanese populations, and Japanese populations  
 412 seem to have originated by a recent host switch from the Taiwanese population to  
 413 the South Clade (Fig. 5). The common ancestor of Clade I was estimated to  
 414 parasitize the South deer clade, and the louse populations on the North deer clade  
 415 seem to have originated by a very recent host switch from louse populations  
 416 parasitizing the South deer clade (Fig. 5).

417       This scenario is reasonable based on the additional evidence provided by  
 418 genetic distances (Figs 3, 6, Suppl. Fig. 2). Deer lice collected from Kyushu  
 419 contain haplotypes from all major louse clades (Clade Ia, Ib and II) and, even within  
 420 Clade I, they are quite variable genetically (shown by dotted circles: Fig. 3). In  
 421 contrast, although lice from Hokkaido and Honshu are also divided into two  
 422 subclades (Clade Ia and Ib), their haplotypes are either identical with (Clade Ia) or  
 423 only slightly different from (Clade Ib) those of Kyushu (Fig. 3). Therefore, recent  
 424 host switches of deer lice from Kyushu to Hokkaido + Honshu populations are  
 425 highly plausible.

426       This scenario implies two possibilities. The first possibility is that the common  
 427 ancestor of the Northern deer clade might not have hosted any louse. The  
 428 Northern deer clade is thought to have colonized the Japanese Archipelago during  
 429 the last ice age via Sakhalin (see below). If deer lice are not tolerant of cold  
 430 temperatures, this scenario might be plausible. Although no quantitative data are  
 431 available, we only collected two lice from two deer at Nishiokoppe, the northern  
 432 most locality of the present sampling area (Fig. 1), and the field work was  
 433 conducted in mid-winter (Table 1). Based on other collections we have made, this  
 434 was an exceptionally small number of lice infecting these deer. A second possibility  
 435 is that the louse populations originally parasitizing deer in Hokkaido and Honshu  
 436 were completely replaced by those from Kyushu following the recent host switch

437 events. Recently, *Damalinia sika* was introduced to the USA has since become  
438 widespread and is replacing native lice very rapidly in some native deer populations  
439 (Mertins *et al.* 2011, unpub. data). This may indicate that certain lineages of deer  
440 lice, particularly parthenogenetic species such as *Damalinia sika*, may have higher  
441 population growth rates, giving them the potential to replace other populations.

442

#### 443 *Implications of Louse Population Structure for Deer Phylogeography*

444 Previous studies from the mitochondrial D-loop and CytB sequences show  
445 that Japanese sika deer possess two genetically distinct lineages (Tamate *et al.*  
446 1998; Nagata *et al.* 1999). Using sequences of the COI gene, the present analysis  
447 also supported this previous result (Fig. 2). However, as discussed above, louse  
448 phylogeography is highly incongruent with deer population structure (Fig. 3), even  
449 though one would expect that dispersal of these permanent parasites is highly tied  
450 to that of their hosts.

451 One possible explanation for the topological incongruence may be phoresis of  
452 these lice on hippoboscids as has been convincingly demonstrated for pigeon  
453 lice (Harbison *et al.* 2009; Harbison and Clayton, 2011). Unlike lice, hippoboscids  
454 flies have wings and can disperse between host individuals relatively freely. Sika  
455 deer host three hippoboscid flies, *Lipoptena fortisetosa*, *L. sika* and *L. japonica*  
456 (Mogi, 1975; Mogi *et al.* 2002). The final instar larvae of these species leave the  
457 deer for pupation and, after emergence, louse-free adult flies fly to new hosts. Soon  
458 after the arrival to new host deer, the flies cut off their wings and thus are not  
459 likely to disperse to a new host (Yamauchi and Nakayama, 2006; Yamauchi,  
460 personal communication). In addition, there are no records of *Damalinia sika*  
461 attached to hippoboscids. Therefore, although the possibility cannot be  
462 excluded completely, phoresis does not seem a likely explanation for the topological  
463 incongruence between deer and louse trees.

464 The more likely explanation for incongruence between deer and louse trees is  
465 that lice migrate via male deer. The two geographically distinct lineages of Japanese  
466 sika deer have only been recovered using maternally inherited mitochondrial



467 markers, which only represents the genetic structure of female deer (Nagata *et al.*  
468 1999). In contrast, two such geographically distinct groups are not recovered in  
469 analyses of microsatellite markers (Goodman *et al.* 2001). In fact, the genetic  
470 structure of Japanese sika deer recovered using microsatellite markers is more  
471 congruent with the louse phylogeny, because the deer populations from Kyushu are  
472 divided into two clusters based on microsatellites. Furthermore, microsatellite data  
473 clusters a deer population from northern Kyushu together with populations from  
474 northern Honshu and Hokkaido (Goodman *et al.* 2001), which is also concordant  
475 with the louse phylogeny (Figs 1 and 3). Sika deer are polygamous, and females  
476 compose maternal groups that do not migrate and tend to stay within a male's  
477 territory. In contrast, young male deer (3–7 year old) migrate widely before  
478 obtaining a harem (Miura, 1984; Takatsuki, 2006). Therefore, deer lice could be  
479 dispersed widely by male deer, and the dispersal of males is likely to be the main  
480 source for the incongruence between deer and lice mitochondrial population  
481 histories. Observations from deer nuclear markers suggest that the louse phylogeny  
482 does at least partially reflect the deer's population structure, with respect to the  
483 fact that male gene flow is quite frequent between two mitochondrial lineages of  
484 sika deer. The louse population structure also provides further support for the idea  
485 that subspecific division in sika deer does not reliably reflect population  
486 subdivision.

487       The two Japanese mitochondrial lineages of sika deer have been estimated to  
488 have already diverged on the continent before their invasion of the Japanese  
489 Archipelago (around 0.3–0.5 Mya), based on mitochondrial genetic distances and  
490 geographic history of the region (Nagata *et al.* 1999; Okada, 2008). The Southern  
491 Clade is hypothesized to have invaded via the Korean Peninsular in the mid  
492 Pleistocene (–0.17 Mya), and the Northern Clade is estimated to have invaded via  
493 Sakhalin in late Pleistocene (60,000–10,000 ya). However, as mentioned above,  
494 these estimates are based on only female line genetic structure. In the present  
495 analysis, the Taiwanese deer louse is placed very close to the deer louse from  
496 southern Kyushu (Fig. 3), which implies that southern Japanese populations of sika

497 deer may have had contact with continental deer more recently than estimated from  
498 the mitochondrial phylogeny of the deer alone.

499 As pointed out in previous studies (Johnson *et al.* 2002a; Clayton and  
500 Johnson, 2003; Weckstein, 2004; Poulin *et al.* 2011), detailed information about  
501 louse and host behaviors is important for understanding the causes of similarities  
502 and discordance between the host and louse phylogenies. The behavior of the sika  
503 deer is well studied (reviewed in Takatsuki, 2006), which greatly aided in  
504 interpreting the discordance between the deer and louse phylogenies. Alternatively,  
505 studies of louse phylogeography could also prove important for understanding the  
506 host's population structure. The substitution rate of louse mitochondrial genes is  
507 much faster than that of their vertebrate hosts (see below), and these insects can  
508 be transmitted both paternally as well as maternally. In the case of sika deer, good  
509 nuclear markers for population genetic studies, including those on Y-chromosome,  
510 have not been discovered to date (Barbosa and Carranza, 2010), and long distance  
511 dispersal of deer lice apart from their hosts is quite unlikely. In such cases, louse  
512 population genetic trees can be good indicators of hidden population structure in  
513 their hosts (Reed *et al.* 2004; Whiteman *et al.* 2007; Zohdy *et al.* 2012). This  
514 notion seems to be supported in our study by the fact that the louse tree is more  
515 similar to the deer population structure estimated from nuclear microsatellite  
516 markers (Goodman *et al.* 2001) than to that estimated from mitochondrial genes  
517 (Nagata *et al.* 1999).

518

#### 519 *Comparisons of Substitution Rates*

520 By plotting pairwise uncorrected genetic distance comparisons for lice against  
521 those for their deer, roughly five groups of points are evident (Fig. 6). Group A  
522 represents the shallowest divergences within both deer and lice. The phylogenetic  
523 trees do not show significant evidence for unevenness of substitution rates among  
524 populations for either deer or lice. Therefore, one of the slopes connecting A-B,  
525 A-E-C or A-D likely estimates the relative rate of substitution for lice vs. deer of  
526 the COI gene. As discussed above, plot groups B and C clearly represent some

527 incongruence between louse and deer population history (Figs 4–6). Given these  
528 considerations, the slope of the line connecting groups of comparisons A and D  
529 seems to be the best estimate for the relative rate of substitution between lice and  
530 deer (Fig. 6). Most of the comparisons between lineages involved in codivergence  
531 (see below and Results) are also in Groups A and D.

532       This slope estimates the relative substitution rate to be around 4.8 times  
533 faster in lice than in deer when uncorrected pairwise distances are used.  
534 Comparisons of four-fold degenerate sites (i.e. neutral sites: 137 bp for deer and  
535 131 bp for lice) also provided a comparable value (4.9 times). This relative rate is  
536 greater than those estimated previously (two to four times faster) for lice and their  
537 hosts (Hafner *et al.* 1994; Page, 1996; Huelsenbeck *et al.* 1997; Page *et al.* 1998;  
538 Reed *et al.* 2004; Light & Hafner, 2007; Demastes *et al.* 2012). Phylogenetic  
539 studies show that mitochondrial substitution rate of deer lice is faster than that of  
540 other lice (Yoshizawa and Johnson, 2003, 2010: note that the species is mislabeled  
541 as *Bovicola tibialis* in Yoshizawa and Johnson, 2003: see Mertins *et al.* 2011 for  
542 further information). Therefore, the presently estimated relative rate (4.8 times)  
543 can be regarded as reasonable. Although a far greater relative rate (11 times) was  
544 estimated by comparing the branch lengths of likelihood trees that were congruent  
545 between doves and their wing lice (Johnson *et al.* 2003b), a smaller estimate of  
546 relative rate of 1.6 times was obtained based on parsimony. Although not shown,  
547 comparisons of likelihood-based pairwise distances for deer and louse COI  
548 sequences resulted in a greater relative substitution rate (7.1 times). This value  
549 does not account for topology because of the many non-codivergence events  
550 identified in the present analyses. Estimates based on branch lengths in a tree  
551 topology probably provide higher relative rates because more multiple substitutions  
552 can be accounted for using a tree based approach, as was done for doves and their  
553 wing lice (Johnson *et al.* 2003b). As mentioned above, the substitution rates of deer  
554 louse mitochondrial genes are faster than other lice sequenced to date (Yoshizawa  
555 and Johnson, 2003, 2010), possibly related to its parthenogenesis (Bengtsson,  
556 2003; Henry *et al.* 2011; Shreve *et al.* 2011).

557

558 *Microevolution of Louse Head Morphology*

559       The dendrogram obtained by cluster analysis of louse head morphology was  
560 highly concordant with the louse phylogenetic tree (Figs 3, 8). The only obvious  
561 discordance was a difference in the placement of populations from Nara and  
562 Kinkazan, otherwise all the major clusters were concordant with the louse  
563 molecular tree. Thus, the shape of the louse head capsule appears to contain  
564 significant phylogenetic signal.

565       The morphology of the conus and neighboring structures shows the greatest  
566 variation and these variables are the major factors in PCs 1–3 and major or minor  
567 factor in PCs 4–6 (Fig. 7). The function of the conus is poorly understood.  
568 However, this structure often shows sexual dimorphism and is frequently absent in  
569 male Trichodectidae (Lyal, 1985) so that the conus may not be a highly functional  
570 structure and thus selective pressure on this character may be relatively weak. The  
571 second structure showing large variation is the anterior margin of the pulvinus (Fig.  
572 7). This character is one of the factors in PCs 1, 2 and the major factor in PC 4.  
573 This structure is involved in grasping the host hair and thus is functionally  
574 important. Previous study has shown that this character is strongly correlated with  
575 host size and thus is likely to be under strong selection pressure (Reed *et al.*  
576 2000).

577       The factors explaining the largest variation in louse head morphology appear  
578 to be both relatively neutral as well as characters expected to be under stronger  
579 selection. Variation in these characters clearly contains signal concordant with the  
580 louse phylogeographic history (Figs 2, 8). In this case, the neutral characters  
581 appeared to comprise a larger fraction of the variation compared to those expected  
582 to be more functionally significant. This suggests that random drift of more neutral  
583 characters plays a major role in the microevolution of morphology for these lice.  
584 The evolution of functionally significant characters is slower compared to that of  
585 less functional characters, perhaps because of stabilizing selection. Interestingly,  
586 the shape of conus is frequently used in taxonomic and phylogenetic studies, but

the shape of pulvinus is rarely used (e.g., Lyal, 1985; Smith, 2001). A high level of variation and correlation with phylogeny of taxonomically important characters, such as dorsal head pattern and abdominal pigmentation, have also been identified in the louse genus *Brueelia* (Johnson *et al.* 2002a), though functional significance of these characters is less clear. Many important diagnostic characters used in louse taxonomy possibly originated by the accumulation of more neutral variation along independent phylogenetic lineages.

The present analysis shows that microevolutionary changes in deer louse morphology accumulate along phylogenetic lines (Figs 3, 8). In contrast, Smith *et al.* (2004) and Johnson *et al.* (in press) showed that, across bird feather lice, convergence is frequent in functionally significant characters, such as body width (i.e., body vs wing lice) or the shape of the anterior head margin. Such convergence is evidence that natural selection produced the variation seen in louse morphology across bird lice. The morphology of deer lice also provides a potential example of convergence caused by similar selection pressures. Although most morphometric clusters are concordant with the louse phylogenetic tree, populations from Kinkazan and Nara were placed at discordant positions (Fig. 8). Interestingly, sika deer of these populations have unusual histories. Kinkazan is a small island (9.6 km<sup>2</sup>), and the population size of sika deer on this island has been highly variable. Given this population fluctuation in their hosts, it is also likely that the lice would have experience population bottlenecks. Dwarfism of deer on Kinkazan also occurs because of the occasional high deer density on the island. Similarly, sika deer in Nara inhabit an urbanized environment (Nara Park), and their population is controlled by humans. Large decreases in population size have occurred in the past. In addition, these deer exhibit dwarfism in body size and delay of sexual maturity (Torii and Tatsuzawa, 2009). These phenomenon are thought to be a result of selection when the deer in Nara Park are at high density. These unusual environmental conditions and/or population bottlenecks probably resulted in accelerated morphological evolution in deer lice that is not concordant with their phylogenetic history.

617

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637

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842  
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844 **Figure legends**

845 Fig. 1. Map of collecting sites for sika deer and lice. Circles indicate the collection  
846 sites, with different colors showing different louse clades. For numbers of deer  
847 and lice samples from each locality, see Table 1.

848 Fig. 2. Phylogenetic tree of sika deer individuals estimated using maximum  
849 likelihood. Branch lengths are proportional to substitutions per site. Numbers  
850 associated with each branch indicate MP bootstrap/ ML bootstrap/ Bayesian  
851 posterior probabilities. Shades indicate haplotype group names used for Figs  
852 4–5. Numbers in circles indicate codivergence points corresponding to those  
853 in Fig. 3. See also Figs 4–6.

854 Fig. 3. Phylogenetic tree of deer lice estimated using maximum likelihood. Branch  
855 lengths are proportional to substitutions per site. Numbers associated with  
856 each branch indicate MP bootstrap/ ML bootstrap/ Bayesian posterior  
857 probabilities. Shades indicate haplotype group names used for Figs 4–5.  
858 Numbers in circles indicate codivergence points corresponding to those in  
859 Fig. 2. Broken-line circles indicate samples from Kyushu and adjunct islands.  
860 See also Figs 4–6.

861 Fig. 4. Tanglegram of deer and louse trees with lines connection host–parasite  
862 associations.

863 Fig. 5. Optimal Jungle reconstruction using TreeMap2. Note that two host switch  
864 events indicated by asterisks were added by hand, because of the inability of  
865 Jungles analysis to deal with parasite lineages occurring on multiple host  
866 lineages. Labels for louse are distinguished from deer by a “p” heading. See  
867 text for detailed explanation.

868 Fig. 6. Plots of uncorrected pairwise distances estimated from deer and louse COI  
869 sequences. Shadings indicate deer and lice divergence events as noted on left  
870 and lower margins. Dashed outlines and associated letters indicate plot  
871 groups as discussed in the text. Numbers in circles indicate codivergence  
872 points corresponding to those in Figs 2–3 and 5.

873 Fig. 7. Illustration of shape variation in louse head morphology, with variation

874 contained within each of the first four principal components indicated.  
875 Fig. 8. Dendrogram resulting from cluster analysis of the louse head shape principal  
876 components. Terminal labels indicate louse collection site (left), their host  
877 subspecies (represented by first three letters of subspecific epithet: middle),  
878 and the mitochondrial lineages for each louse population (right).

879

880 **Table 1.** List of the samples used in this study. Abbreviated labels corresponding to  
881 those indicated in figures.

882

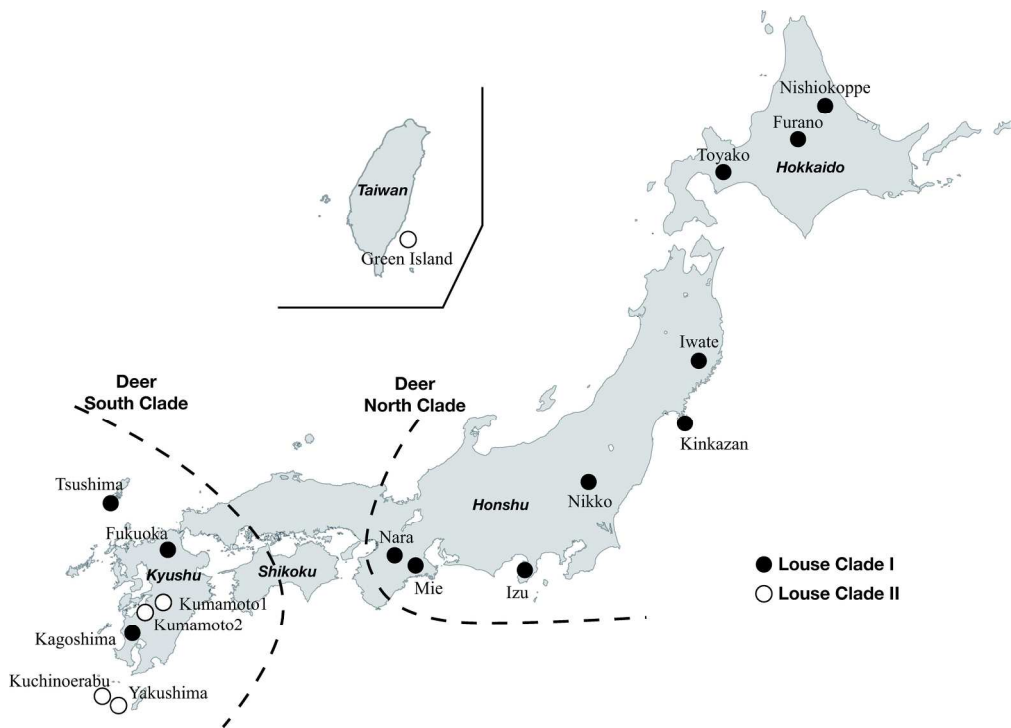
883 **Supplementary Figure 1.** ParsimonySplits network of sika deer.

884

885 **Supplementary Figure 2.** ParsimonySplits network of sika deer lice.

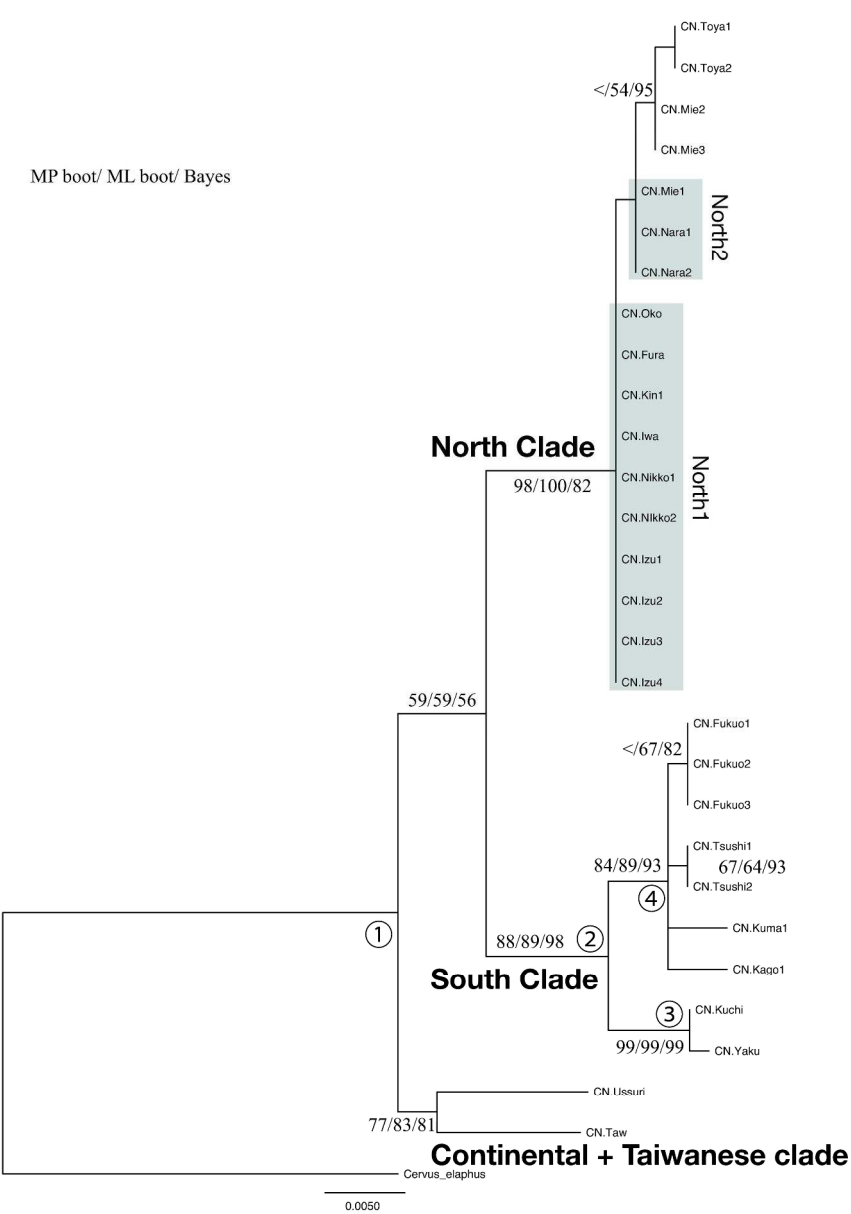
886

887 **Supplementary data.** Primer list and selected evolutionary models for likelihood and  
888 Bayesian analyses (data.txt), raw data from Fourier analyses (sape.data.csv),  
889 aligned sequences (Lice.coi.full.nex: full data for lice; Deer.coi.full.nex: full  
890 data for deer; Louse-deer.coi.nex: combined 1:1 data set), and deer ML tree  
891 estimated without a sample from Ussuri (exUssuri.tre).

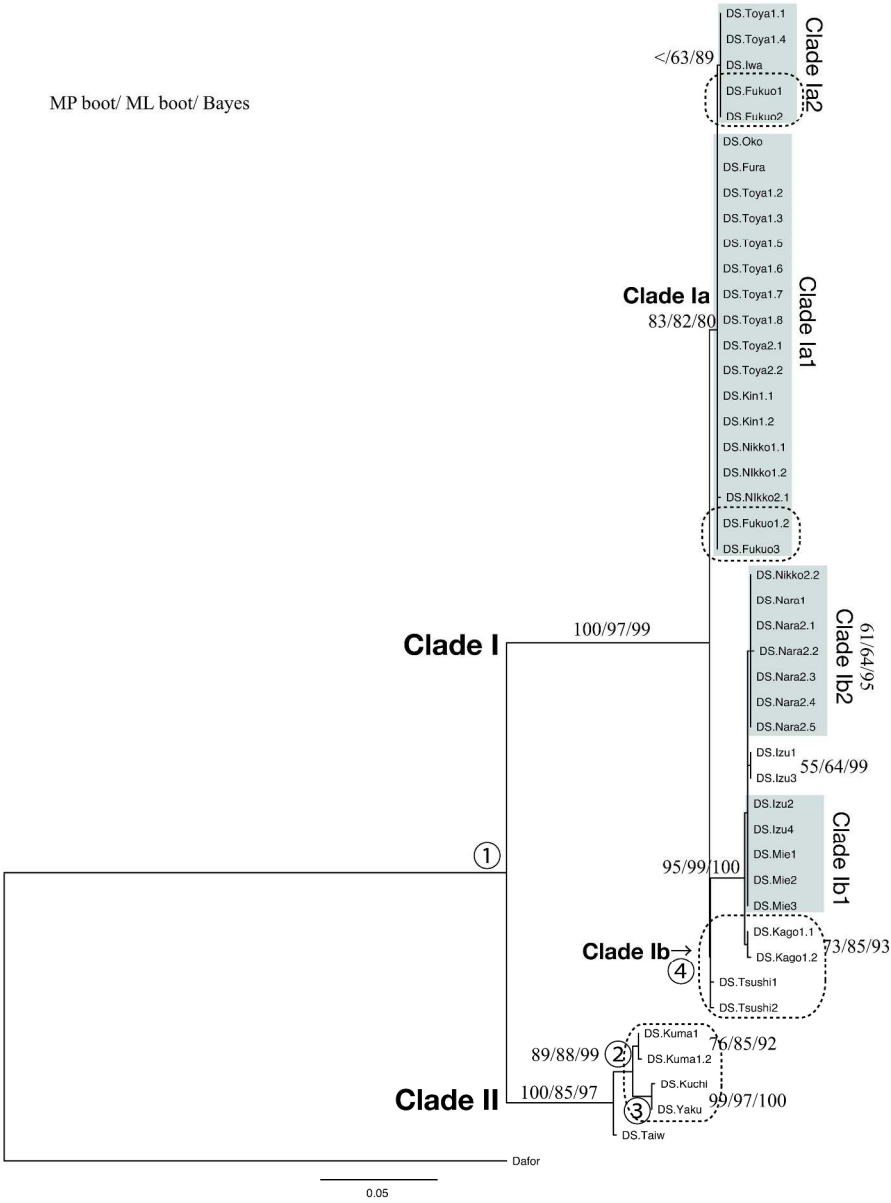


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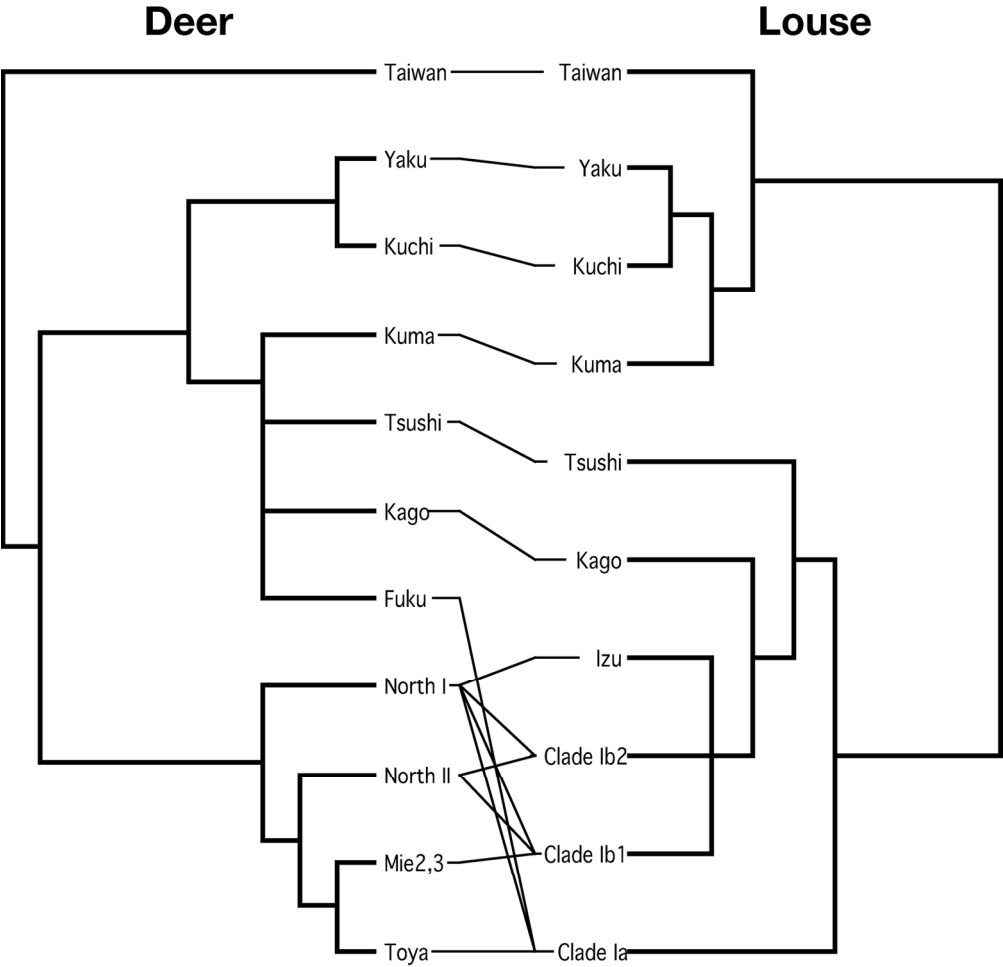




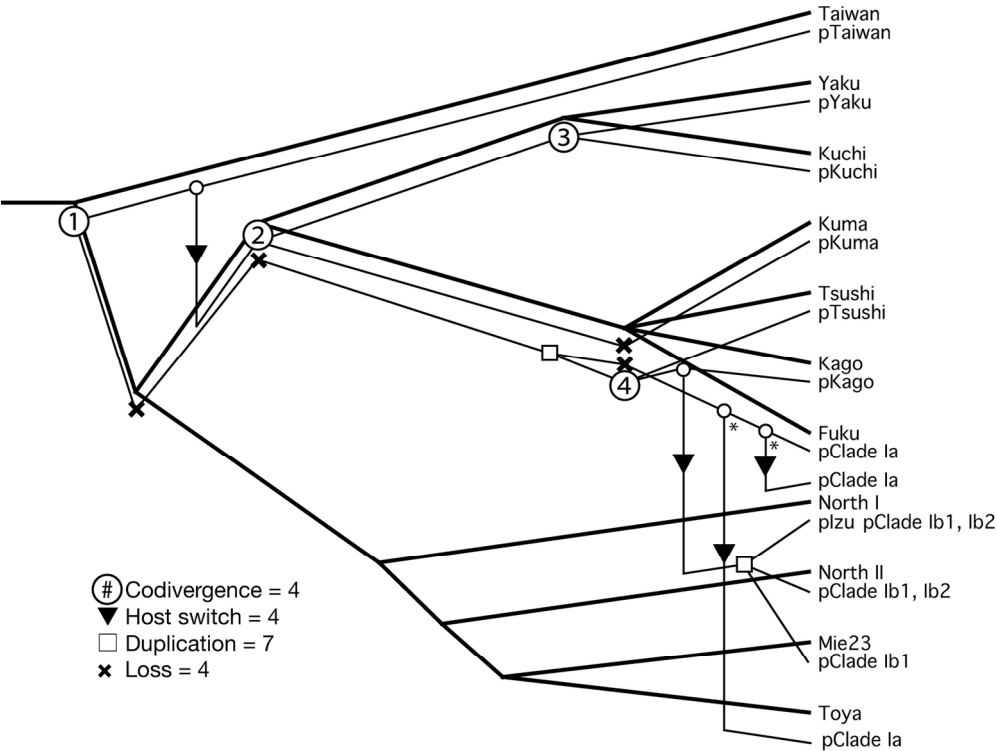
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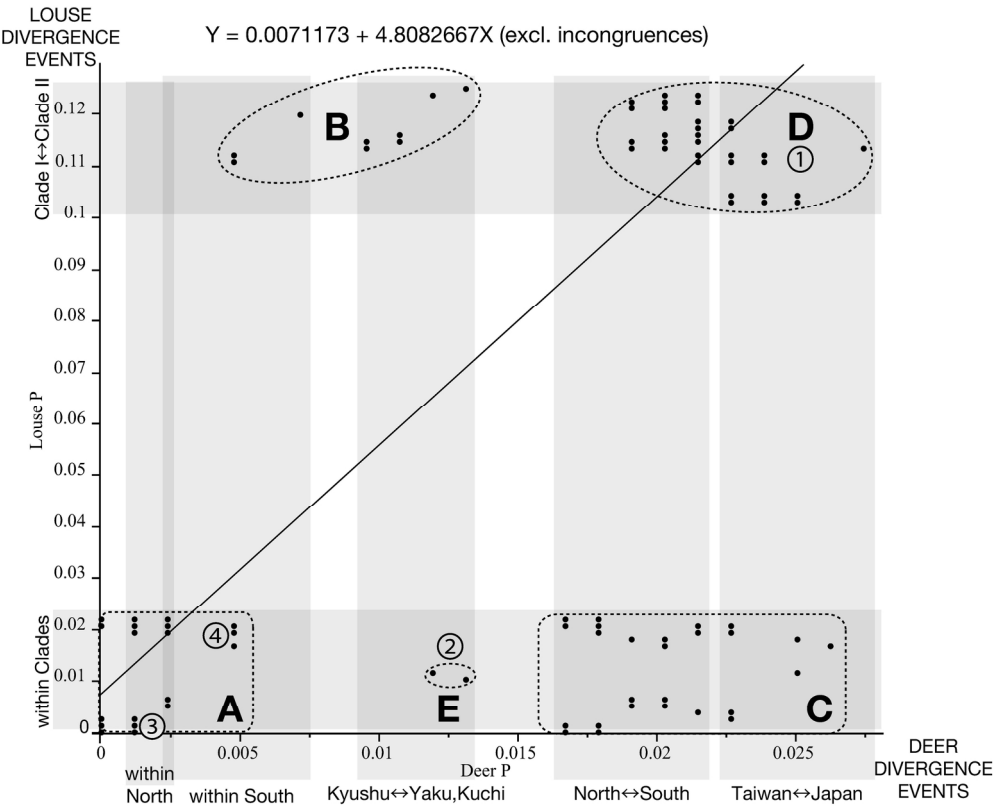
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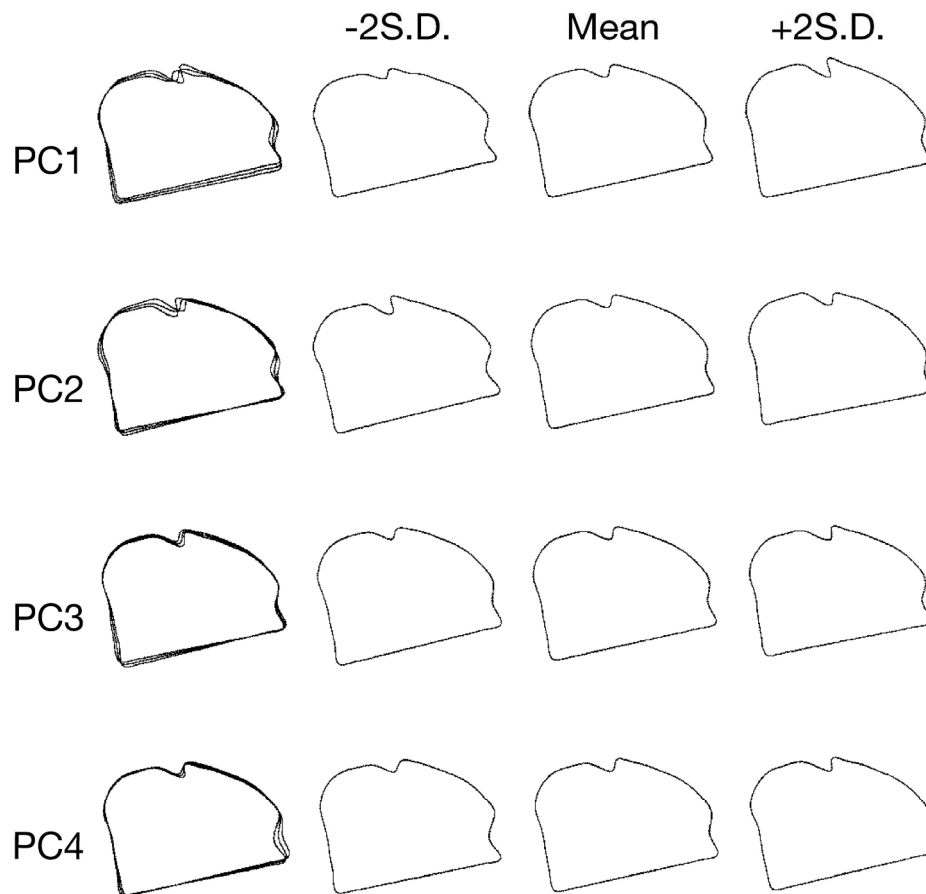
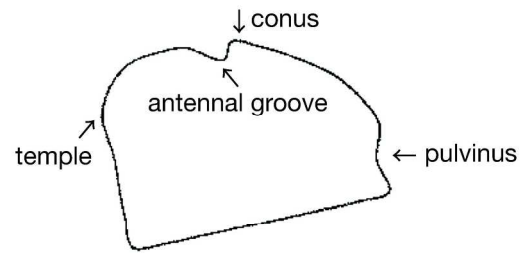
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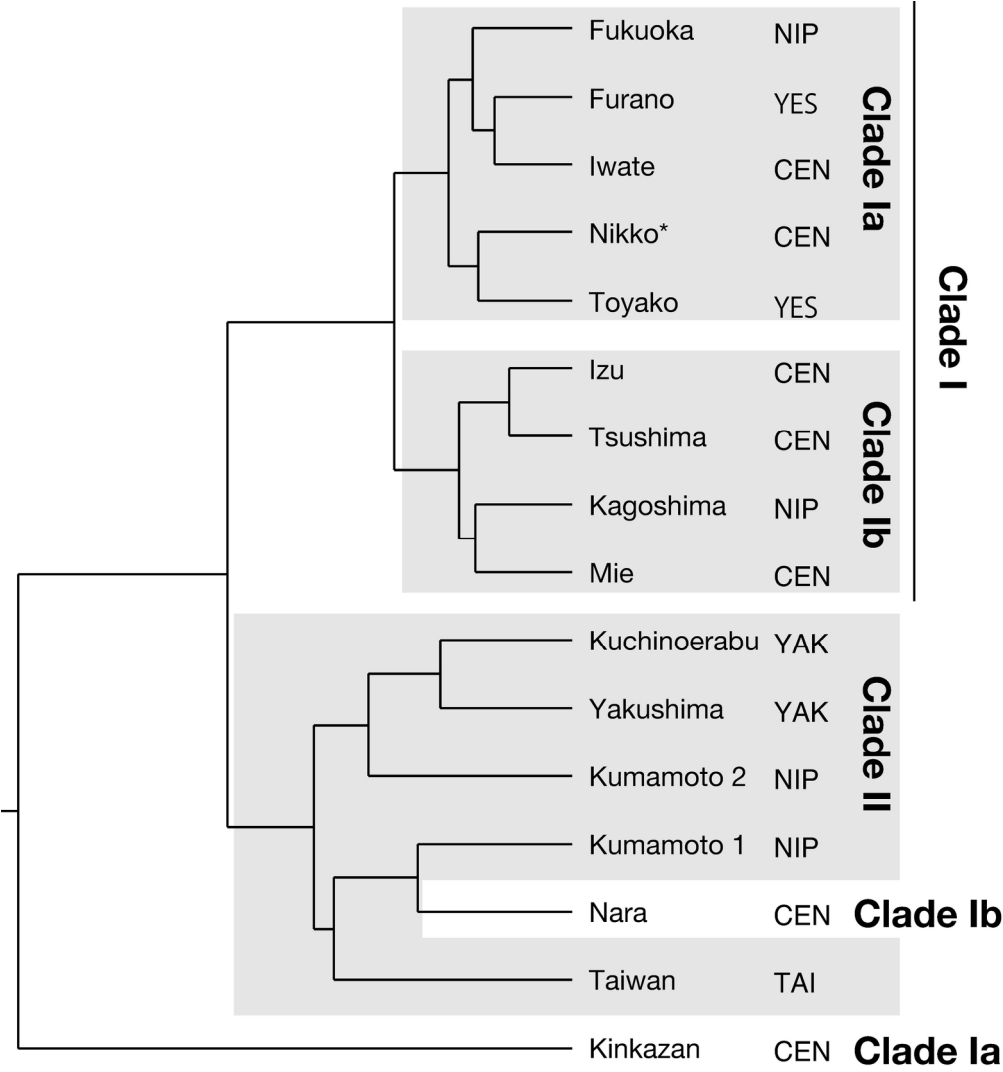
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179x143mm (300 x 300 DPI)



202x261mm (300 x 300 DPI)



179x192mm (300 x 300 DPI)

Label	Deer subsp.	Locality	Date	DNA (n)		Morph (n)	GenBank	
				Deer	Louse	Louse	Deer	Louse
<b>Oko</b>	<i>yesoensis</i>	Hokkaido: Nishiokoppe Villa.	20. ii. 2007	1	1	n.a.	AB713061	AB713088
<b>Fura</b>	<i>yesoensis</i>	Hokkaido: Furano City, Experimental Forest of the Univ. Tokyo	23. xii. 2007	1	1	10	AB713062	AB713089
<b>Toya</b>	<i>yesoensis</i>	Hokkaido: Lake Toyako, Nakajima	7. iii. 2002	2	10	10	AB713063-4	AB713090-9
<b>Iwa</b>	<i>centralis</i>	Iwate: Sumita Town	18. i. 2004	1	1	4	AB713065	AB713100
<b>Kin</b>	<i>centralis</i>	Miyagi: Kinkazan Is.	4. vi. 2001	1	2	2	AB713066	AB713101-2
<b>Nikko</b>	<i>centralis</i>	Tochigi: Nikko City	30. i and 26. ii. 2009	2	3	10	AB713067-8	AB713103-6
<b>Izu</b>	<i>centralis</i>	Shizuoka: Izu City, Mt. Tanabayama	4. iii. 2009	4	4	10	AB713069-72	AB713107-10
<b>Mie</b>	<i>centralis</i>	Mie: Odai Town	ii. 2009	3	3	10	AB713073-5	AB713111-3
<b>Nara</b>	<i>centralis</i>	Nara: Nara Park	1. v. 2008	2	6	10	AB713076-7	AB713114-9
<b>Tsushi</b>	<i>centralis</i>	Nagasaki: Tsushima Is., Izuhara Town	28. i and 8. ii. 2009	2	2	10	AB713078-9	AB713120-1
<b>Fuk</b>	<i>nippon</i>	Fukuoka: Asakura C.	17 and 24. i. 2009	3	4	10	AB713080-2	AB713122-5
<b>Kuma1</b>	<i>nippon</i>	Kumamoto: Izumi Villa., Shiibagoe	24. v. 1997	1	1	10	AB713083	AB713126
<b>Kuma2</b>	<i>nippon</i>	Kumamoto: Kuma Villa.	7. iv. 2010	n.a.	1	8	n.a.	AB713127



Kago	<i>nippon</i>	Kagoshima: Satsuma Town, Mt. Shibaoyama	10 and 14, iii. 2008	1	2	10	AB713084	AB713128-9
Kuchi	<i>yakushimae</i>	Kagoshima: Is. Kuchinoerabu	25. ii. 2008	1	1	5	AB713085	AB713130
Yaku	<i>yakushimae</i>	Kagoshima: Is. Yakushima, Miyanoura	3. iii. 2008	1	1	10	AB713086	AB713131
Taiw	<i>taiouanus</i>	Green Island, Taiwan	19. ix. 2006	1	1	2	EF058308	AB713132
Ussuri	<i>hortulorum</i>	GenBank	-	1	n.a.	n.a.	NC013834	n.a.
Dafor (louse outgroup)	-	Marin County, California, USA ex. Axis axis	22. vi. 2005	n.a.	1	n.a.	n.a.	AB713087
Cervul elaphus (deer outgroup)	-	GenBank	-	1	n.a.	n.a.	AB245427	n.a.

Review

[supplementary data file for co-phylogeography of sika deer and deer lice:  
Mizukoshi, Johnson and Yoshizawa]

PCR and Sequencing primers for deer

COI.CN.F (GCA GCC GGA ATT ACA ATA CT) + COI.CN.R2 (GCA ACT ACA TAA TAT GTG  
TCA)

COI.CN.F2 (CTA GCA ACA CTC CAC GGA GG) + COI.CN.R (GCA TCC ATT TAG YCA CTC TA)

PCR and Sequencing primers for lice

COI.DS.F (ATY ACA ATG CTT CTT CTA GAT CG) + COI.DS.R2 (TGG AAR TGY GCT ACC ACR  
TA)

COI.DS.F2 (CAC CAT TGG RGG TCT CAC TG) + COI.DS.R (AAT TCG ATA AGA GAA CAA GGA  
GC)

Likelihood model selected for lice data

[!Likelihood settings from best-fit model (HKY+G) selected by hLRT in  
Modeltest 3.7 on Fri Oct 7 10:25:20 2011]

BEGIN PAUP;

Lset Base=(0.2101 0.1952 0.2412) Nst=2 TRatio=4.2614 Rates=gamma

Shape=0.3251 Pinvar=0;

END;

Likelihood model selected for deer data

[!Likelihood settings from best-fit model (HKY+G) selected by hLRT in  
Modeltest 3.7 on Fri Oct 7 10:23:26 2011]

BEGIN PAUP;

Lset Base=(0.2935 0.2196 0.1632) Nst=2 TRatio=22.5927 Rates=gamma

Shape=0.0106 Pinvar=0;

END;

Bayesian model selected for lice data

[!MrBayes settings for the best-fit model (HKY+G) selected by hLRT in  
MrModeltest 2.3]

BEGIN MRBAYES;

Lset nst=2 rates=gamma;

Prset statefreqpr=dirichlet(1,1,1,1);

END;

Bayesian model selected for deer data

[!MrBayes settings for the best-fit model (HKY+G) selected by hLRT in  
MrModeltest 2.3]

BEGIN MRBAYES;

Lset nst=2 rates=gamma;

Prset statefreqpr=dirichlet(1,1,1,1);

END;

