1 Submitted to Applied and Environmental Microbiology $\mathbf{2}$ Symbiotic bacteria associated with stomach disc of human lice 3 4 Kayoko Sasaki-Fukatsu,¹ Ryuichi Koga,¹ Naruo Nikoh,² Kazunori Yoshizawa,³ Shinji Kasai,⁴ $\mathbf{5}$ Minoru Mihara,⁵ Mutsuo Kobavashi,⁴ Takashi Tomita,⁴ and Takema Fukatsu¹* 6 78 Institute for Biological Resources and Functions, National Institute of Advanced Industrial Science and Technology (AIST), Tsukuba 305-8566,¹ Division of Natural Sciences, University 9 of the Air, Chiba 261-8586,² Department of Ecology and Systematics, Hokkaido University, 10 Sapporo 060-8589,³ Department of Medical Entomology, National Institute of Infectious 11 Diseases, Tokyo 162-8640,⁴ and Department of Environmental Biology, Japan Environmental 12Sanitation Center, Kawasaki 210-0828,⁵ Japan 1314* Corresponding author. Mailing address: National Institute of Advanced Industrial Science 15and Technology (AIST), 1-1-1 Higashi, Tsukuba 305-8566, Japan. Phone: 81-29-861-6087. 16 Fax: 81-29-861-6080. E-mail: t-fukatsu@aist.go.jp. 1718 19 Running head: Symbiotic bacteria of human lice 20

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22The symbiotic bacteria associated with stomach disc, a large aggregate of bacteriocytes on the 23ventral side of midgut, of the human body and head lice were characterized. Molecular 24phylogenetic analysis of 16S rRNA gene sequences showed that the symbionts formed a 25distinct and well-defined clade in the γ -Proteobacteria. The sequences exhibited AT-biased 26nucleotide composition and accelerated molecular evolution. In situ hybridization revealed 27that in nymphs and adult males the symbiont was localized in the stomach disc, while in adult 28females the symbiont was not in the stomach disc but in the lateral oviducts and the posterior 29pole of the oocytes due to female-specific symbiont migration. We propose the designation 30 "Candidatus Riesia pediculicola" for the lice symbionts.

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32Sucking lice (Insecta: Phthiraptera: Anoplura), embracing over 500 described species 33 from the world, are ectoparasitic insects exclusively feeding on mammalian blood (9). There 34are two closely related species of human lice: the body louse *Pediculus humanus* lives in 35clothes and feeds from body, and the head louse *Pediculus capitis* lives in hair and feeds from 36 scalp. The head louse and the body louse are morphologically and genetically very similar, 37 and some researchers regard them as subspecies or ecotypes of the same species (7, 16, 22).

38 Vertebrate blood is certainly nutritious but deficient in some nutritional components such 39 as B vitamins, which is probably the reason why insects exclusively living on vertebrate 40 blood all through their life, including tsetse flies, louse flies, bedbugs, assassin bugs, lice, etc., 41 are generally in close association with endosymbiotic microorganisms (6). It was back to the 421920's when the human lice were first reported to possess a large aggregate of bacteriocytes, 43called stomach disc, on the ventral side of midgut, wherein rod-shaped symbiotic bacteria are 44harbored (5, 23). Since then, a number of histological (11, 20), embryonic (4, 20), 45experimental (1, 2, 3, 10) and nutritional (18, 19) studies have been conducted on the endosymbiotic system of the human lice. These studies demonstrated that the symbiont is 46 47vertically transmitted from maternal stomach disc to developing oocytes through a peculiar passage (11, 20), is essential for survival and growth of the host (3), and provides the host 48 with B vitamins that are lacking in the blood meal (19). Despite the substantial body of early 4950works on the lice endosymbiosis, microbial nature of the symbionts has been unknown to date. 51Hence, we characterized the symbiotic bacteria of the human lice by using molecular 52phylogenetic and histological approaches.

53We mainly used a long-established inbred line of the human body louse, the strain NIID, 54which has been maintained in the laboratory since 1954 (26). The insects were kept in an 55evaporating dish with pieces of felt sealed in a plastic container with silica gel, reared at 30°C 56in constant darkness, and fed with human blood once a day on the arm of one of the authors 57(M. M.). The insects at different developmental stages were sampled and preserved in acetone 58until molecular and histological analyses (13). Samples of body lice from 12 different sources 59and head lice from 4 different sources were also collected in either Japan or Nepal.

60 DNA was individually extracted from adult females of the strain NIID by using a 61 QIAamp DNA mini kit (QIAGEN). A 1.5 kb segment of the eubacterial 16S rRNA gene was 62 amplified with the primers 16SA1 (5'-AGAGTTTGATCMTGGCTCAG-3') and 16SB1 63 (5'-TACGGYTACCTTGTTACGACTT-3'), and subjected to cloning, restriction fragment 64 length polymorphism (RFLP) genotyping, and DNA sequencing as previously described (14). 65 More than ten clones of the 16S rRNA gene segment from each of the samples exhibited

66 identical RFLP patterns (data not shown), suggesting that a single bacterial species is dominant in the insects. Nucleotide sequences of some of these clones were determined, which were 1,482 bp in size and identical to each other. A BLAST search clearly showed that the sequence belongs to the Enterobacteriaceae in the γ -*Proteobacteria*. No closely related sequences were identified in the DNA databases: the highest hits were *Providencia stuartii* (AF008581; 89.6% [1,141/1,274] sequence identity) and *Enterobacter hormaechei* (AJ853889; 89.0% [1,119/1,258] sequence identity).

73 In order to confirm whether or not the 16S rRNA gene sequence is derived from the 74symbiotic bacteria in the stomach disc, we designed an oligonucleotide probe specific to the sequence, Cy3-Lice1255R (5'-Cy3-TTGGCTCGCTCTTACGAGT-3'), for whole-mount 7576 fluorescent in situ hybridization (wFISH). The insects preserved in acetone were, after their 77 legs were removed to facilitate infiltration of reagents, fixed in Carnoy's solution 78(chloroform-ethanol-acetic acid [6:3:1]) overnight, and incubated with 6% H₂O₂ overnight for 79 quenching the autofluorescence of insect tissues. The insects were thoroughly washed, and 80 equilibrated with a hybridization buffer (20 mM Tris-HCl [pH 8.0], 0.9 M NaCl, 0.01% sodium dodecyl sulfate, 30% formamide), and Cy3-Lice1255R probe and SYTOX green were 81 82 added at final concentrations of 10 nM and 0.5 µM, respectively. After an overnight 83 incubation, the samples were thoroughly washed in a washing buffer (20 mM Tris-HCl [pH 8.0], 0.9 M NaCl, 0.01% sodium dodecyl sulfate), and were observed under an epifluorescent 84 85 microscope (Axiophot, Carl Zeiss) and a laser confocal microscope (PASCAL5; Carl Zeiss).

86 In nymphal insects, the probe specifically hybridized with a round stomach disc located 87 in the ventral abdomen (Fig. 1A). In the stomach disc, the signals showed radial 88 compartment-like structures (Fig. 1B), which had been described as location of the symbiotic 89 bacteria (4, 11, 20). A 3-dimensional FISH movie image of the stomach disc is available in 90 Supplemental Material. No probe control experiment and competitive suppression control 91 experiment with excess unlabelled Lice1255R probe (14) did not identify these signals, 92confirming specificity of the FISH detection (data not shown). These results indicated that the 93 16S rRNA gene sequence was certainly derived from the symbiotic bacteria harbored in the 94stomach disc.

In adult males, the signals were localized in the stomach disc (Fig. 1C). In adult females, however, the signals were not detected in the stomach disc, but in the lateral oviducts and the posterior pole of oocytes in ovarioles (Fig. 1D-F). Ries (20) and Eberle and McLean (11) reported that in the last (= 3rd) nymphal instar, the symbiont cells escape from the stomach disc, migrate to the lateral oviducts, infect the ovarial epithelium, and are transmitted to the oocytes through a specialized tissue organization called ovarial ampullae. The results of wFISH were in accordance with these histological descriptions.

102 We also cloned and sequenced the eubacterial 16S rRNA gene segment from four body 103 lice and two head lice of different sources. All the sequences were very similar to each other, 104 1,482 bp in size, with sequence identities ranging from 99.1% to 99.9%. Nucleotide 105composition of the sequences was remarkably AT-biased, ranging from 50.6% to 51.0%. 106 These AT-content values were higher than those of related free-living bacteria such as P. 107 stuartii (AF008581; 46.9%) and E. hormaechei (AJ853889; 44.9%), and were equivalent to 108those of obligate endosymbiotic bacteria of other insects such as *Buchnera* spp. of aphids (e.g. 109 AJ296751; 51.3%) and Wigglesworthia spp. of tsetse flies (e.g. AB063521; 51.5%).

110 These 16S rRNA gene sequences were subjected to molecular phylogenetic analysis 111 together with the sequences of related γ -proteobacteria that exhibited high BLAST scores in 112 the DNA database search. A multiple alignment of the sequences was generated by using the 113 program package Clustal W (24). Aligned nucleotide sites containing a gap were removed 114 from the data set, and the final alignment was inspected and corrected manually. A 115 neighbor-joining tree, with 1,000 bootstrap resamplings, was also constructed by the program 116 package Clustal W (24).

Figure 2 shows the neighbor-joining phylogeny on the basis of 1,408 unambiguously aligned nucleotide sites. The lice symbionts formed a distinct monophyletic group with 100% bootstrap support, constituting a basal lineage in the γ -subclass of the *Proteobacteria*. The symbionts of body lice could not be phylogenetically differentiated from the symbionts of head lice, probably reflecting the close morphological and genetic relatedness between the body louse and the head louse (7, 16, 22). No endosymbiotic bacteria of other insects showed phylogenetic affinity to the lice symbionts.

124In comparison with the related free-living bacteria, the lice symbionts exhibited 125remarkably elongated branches on the phylogenetic tree (Fig. 2), which was suggestive of 126accelerated molecular evolution in the lineage of the lice symbionts. Thus, we performed a 127relative rate test based on genetic distances estimated under the Kimura's two parameter 128model (15). Statistical significance was evaluated by using the program package RRTree (21). 129The following 16S rRNA gene sequences were subjected to the analysis: AB263101 from 130 symbiont of body louse, AB263105 from symbiont of head louse, AF008581 from 131 Providencia stuartii, AM040495 from Providencia sp., and X74694 from Vibrio cholerae as 132outgroup. The evolutionary rate of 16S rRNA gene in the lineage of the lice symbionts was 133 around 3.1 times faster than that in the lineage of related free-living γ -proteobacteria. The 134difference was highly significant statistically ($P = 10^{-7}$).

Recent molecular evolutionary analyses revealed that the lifestyle of obligate insect endosymbionts has strongly affected their genome evolution, causing AT-biased nucleotide 137 composition, accelerated rate of molecular evolution, and significant genome reduction. 138 These peculiar genetic traits are hypothesized to be the consequence of attenuated purifying 139 selection due to small population size and strong bottleneck associated with their 140 endosymbiotic lifestyle (17, 25). The AT-bias and the accelerated evolution in the 16S rRNA 141 gene sequences are also suggestive of a stable and intimate host-symbiont association in the 142 human lice over evolutionary time.

143How prevalent is the symbiont in the lice populations? We examined 57 body lice from 14412 different sources and 9 head lice from 4 sources for their symbiont infection. For 145confirmation of successful DNA preparation from each of the samples, mitochondrial 16S 146 host insect was amplified with rRNA gene of the the primers MtrA1 147(5'-AAWAAACTAGGATTAGATACCCTA-3') and MtrB3 148(5'-ACACTTTCCAGTACAYTTACTTTGT-3') under a temperature profile of 95°C for 4 149min followed by 35 cycles of 95°C for 30 sec, 48°C for 30 sec and 65°C for 3 min. 150Diagnostic PCR of the symbiont was performed with the primers 16SA3 (5'-TGCATGGYTGTCGTCAGCTCG -3') and Lice1255R under a temperature profile of 15195°C for 4 min followed by 35 cycles of 95°C for 30 sec, 50°C for 30 sec and 70°C for 1 min. 152153The diagnostic PCR analysis identified the symbiont in all the lice samples examined. Thus, it 154was strongly suggested that the symbiont infection is fixed in the lice populations, 155corroborating the essential biological roles of the symbiont for the host insect (3, 19).

156 From all these results taken together, it was concluded that the symbiotic bacteria 157 associated with the stomach disc of the human lice constitute a phylogenetically distinct and 158 coherent bacterial group in the γ -*Proteobacteria*. Hence, we propose the designation 159 "*Candidatus* Riesia pediculicola" for the symbiont of the human lice. The generic name 160 honors Erich Ries who first comprehensively investigated the endosymbiotic system in lice 161 (20). The specific name indicates the association with lice.

162 The human lice are notorious as medical and hygienic pests. The body louse is the vector 163 of the epidemic typhus pathogen Rickettsia prowasekii, the relapsing fever pathogen Borrelia 164recurrentis, and the trench fever pathogen Bartonella quintana (12). The head louse is 165recently re-emerging even in advanced countries, particularly among school children, and is 166 increasing its resistance to insecticides (8). Considering the essential roles of the symbiont for 167the host (3, 19), microbiological studies on the lice symbionts would lead to new means of 168control for the pests. Other than the human lice, a number of lice are known from a wide 169 variety of mammals, and it was histologically described that the localization, morphology and 170 life cycle of symbiotic bacteria in animal lice are distinct from those of the human lice in 171many respects (6, 20). Whether the symbionts of animal lice are phylogenetically related to

172the symbionts of the human lice is of evolutionary interest. 173 174We thank Noboru Yaguchi, Naomi Seki, Madan Shrestha, Harufumi Yui and Akio 175Kobayashi for lice samples. 176 177REFERENCES 1781. Aschner, M. 1932. Experimentelle Untersuchungen über die Symbiose der Kleiderlaus. 179Naturwissenscheften 27:501-505. 180 2. Aschner, M. 1934. Studies on the symbiosis of the body louse 1: Elimination of the 181 symbionts by centrifugation of the eggs. Parasitology 26: 309-314-182 3. Aschner, M., and E. Ries. 1933. Das Verhalten der Kleiderlaus beim Ausschalten der 183 Symbionten. Z. Morphol. Ökol. Tiere 26:529-590. 1844. Baudisch, K. 1958. Beiträge zur Zytologie und Embryologie einiger Insektensymbiosen. Z. 185Morphol. Ökol. Tiere **47**:436-488. 186 5. Buchner, P. 1920. Zur Kenntnis der Symbiose niederer pflanzlicher Organismen mit 187 Pedikuliden. Biol. Zbl. 39: 535-540. 188 6. Buchner, P. 1965. Endosymbiosis of animals with plant microorganisms. Interscience, 189 New York, N.Y. 7. Busvine, J. R. 1978. Evidence from double infestations for the specific status of human 190191 head and body lice (Anoplura). Syst. Entomol. 3:1-8. 192 8. Downs, A. M. R. 2004. Managing head lice in an era of increasing resistance to 193 insecticides. Am. J. Clin. Dermatol. 5:169-177. 194 9. Durden, L. A., and G. G. Musser. 1994. The sucking lice (Insecta: Anoplura) of the world: 195A taxonomic checklist with records of mammalian hosts and geographical distributions. Bull. Am. Mus. Natl. Hist. 218:1-90. 196 197 10. Eberle, M. W., and D. L. McLean. 1982. Initiation and orientation of the symbiote 198 migration in the human body louse *Pediculus humanus* L. J. Insect Physiol. 28:417-422. 199 11. Eberle, M. W., and D. L. McLean. 1983. Observation of symbiote migration in human 200 body lice with scanning and transmission electron microscopy. Can. J. Microbiol. 29:755-762. 20112. Fournier, P.-E., J.-B. Ndihokubwayo, J. Guidran, P. J. Kelly, and D. Raoult. 2002. 202Human pathogens in body and head lice. Emerg. Infect. Dis. 8:1515-1518. 203 13. Fukatsu, T. 1999. Acetone preservation: a practical technique for molecular analysis. Mol. 204 Ecol. 8:1935-1945. 20514. Fukatsu, T., and N. Nikoh. 1998. Two intracellular symbiotic bacteria of the mulberry 206psyllid Anomoneura mori (Insecta, Homoptera). Appl. Environ. Microbiol. 64:3599-3606.

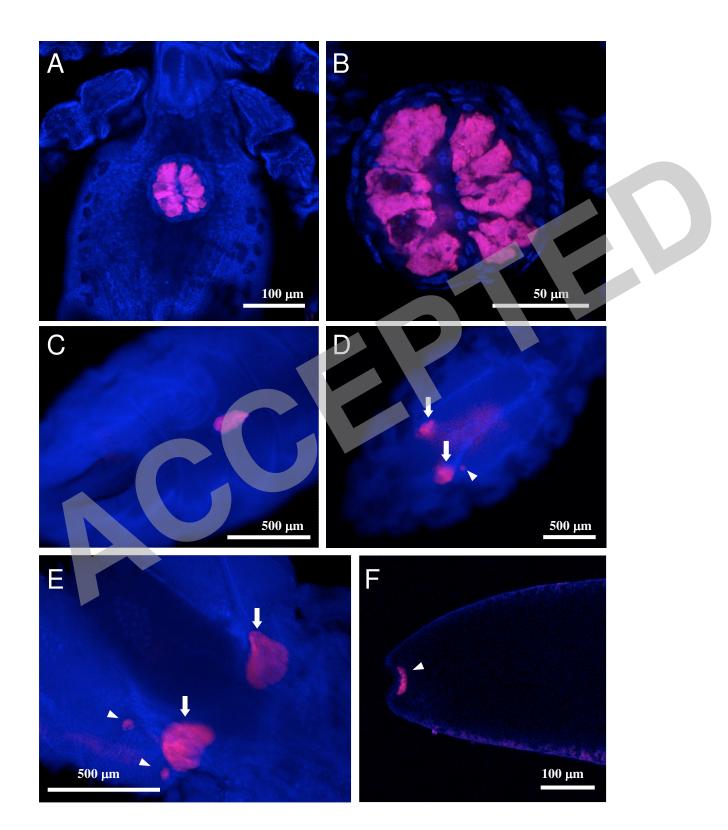
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FIGURE CAPTIONS

- FIG. 1. Whole mount in situ hybridization of the symbiotic bacteria in the human body lice.
- 236 (A) A 1st instar nymph with a large round stomach disc at the center of the ventral abdomen.
- 237 The symbiont cells are specifically detected in the stomach disc. (B) An enlarged image of the
- stomach disc. The location of the symbiont cells exhibited radial compartment-like structures.
- 239 (C) An adult male, in which the symbiont is localized in the stomach disc. (D) An adult
- 240 female, in which the symbiont is not detected in the stomach disc but in the lateral oviducts
- and the posterior pole of oocytes. (E) An enlarged image of the female reproductive organs.

- (F) An enlarged image of an oocyte. Arrows and arrowheads indicate the lateral oviducts andthe posterior pole of oocytes, respectively.
- 244
- FIG. 2. Molecular phylogenetic analysis of 16S rRNA gene sequences of the stomach disc
- 246 symbionts from human lice. A neighbor-joining tree is shown, while maximum parsimony
- analysis gave essentially the same result (data not shown). Bootstrap values higher than 70%
- are shown at the nodes. In brackets are sequence accession numbers.
- 249



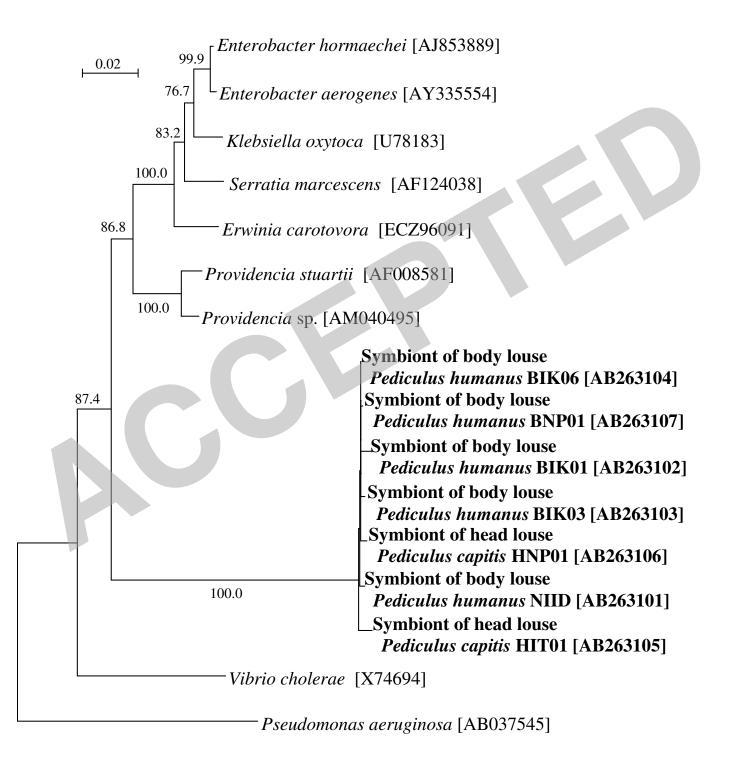


FIG. 2